

Supplementary Information for

The crystal structures of *Arabidopsis* and *Chlamydomonas* phosphoribulokinase fill the last gap in the redox structural proteome of the Calvin-Benson cycle

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References for SI reference citations

Supplementary Information

SI Materials and Methods

Activity Assay and Determination of pH and Temperature Dependence

Phosphoribulokinase activity was measured spectrophotometrically (Cary 60 UV-Vis; Agilent) at 25°C in 1 ml cuvette containing 50 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, 20 mM KCl, 2 mM ATP, 2.5 mM phosphoenolpyruvate, 0.2 mM NADH, 5 U ml⁻¹ pyruvate kinase, 6 U ml⁻¹ lactate dehydrogenase and enzyme preparation. Following 1 min of blank acquisition, the reaction was started by the addition 0.5 mM ribulose-5-phosphate.

Temperature dependence was evaluated on aliquots of *At*PRK and *Cr*PRK desalted in PBS buffer and incubated for 20 min at temperatures ranging from 20°C to 60°C. Following incubation, the activity was assayed at 25°C as described above. Activities were measured on two independent protein purifications and are expressed as percentage of the highest measured activity.

The pH dependence of purified enzymes was determined in Britton-Robinson buffer for pH values ranging from 5.0 to 8.5. For pH values ranging from 9.0 to 10.0, 100 mM Glycine buffer was used. To ascertain that pH and buffer composition do not block pyruvate kinase and lactate dehydrogenase activity, their activity was controlled measuring the rate of NADH oxidation in a coupling assay performed at 25°C in 1 ml cuvette containing the above mentioned buffers and 10 mM MgCl₂, 20 mM KCl, 2 mM ADP, 0.2 mM NADH, 10 U ml⁻¹ pyruvate kinase, 12 U ml⁻¹ lactate dehydrogenase. Following 1 min of blank acquisition, the reaction was started by the addition 2.5 mM phosphoenolpyruvate and monitored spectrophotometrically at 340 nm.

Once the functionality of pyruvate kinase and lactate dehydrogenase was verified, to ascertain that their activity did not limit the measurements of PRK activity, the concentration of both enzymes was increased (i.e. from 5 to 15 U ml⁻¹ for pyruvate kinase and 6 to 18 U ml⁻¹ for lactate dehydrogenase) up to concentrations that did not increase PRK activity further.

Determined the experimental conditions, PRK activity was measured on a Cary 60 UV-Vis spectrophotometer (Agilent) at 25°C in 1 ml cuvette containing the above mentioned buffer at

different pH values, 10 mM MgCl₂, 20 mM KCl, 2 mM ATP, 2.5 mM phosphoenolpyruvate, 0.2 mM NADH, 10 U ml⁻¹ pyruvate kinase, 12 U ml⁻¹ lactate dehydrogenase and enzyme preparation. Following 1 min of blank acquisition, the reaction was started by the addition 0.5 mM ribulose-5-phosphate. Activities were measured on two independent protein purifications and are expressed as percentage of the highest measured activity.

Crystallization and Data Collection.

Aliquots of 2 µl for both proteins were mixed to an equal volume of reservoir, and the prepared drop was equilibrated against 900 (*CrPRK*) and 750 (*AtPRK*) µl of reservoir.

CrPRK crystals were obtained in different conditions of the Extension Kit from Hampton Research (solutions 22: 12% w/v PEG 20K, 0.1 M MES, pH 6.5; 26: 30% w/v PEG MME 5K, 0.1 M MES, pH 6.5, 0.2 M ammonium sulfate; 30: 10% w/v PEG 6K, 5% v/v MPD, 0.1 M HEPES, pH 7.5) and Structure screen 1 from Molecular Dimension (MD1-01-CF, solutions 35: 30% w/v PEG 4K, 0.1 M Tris-HCl, pH 8.5, 0.2 M lithium sulfate; and 50: 15% w/v PEG 8K, 0.5 M lithium sulfate). The conditions were optimized and the best diffracting crystal grew in about 10 days, from 22% w/v PEG MME 5K, 0.1 M MES, pH 6.5, 0.2 M ammonium sulfate. *CrPRK* crystal appeared as needle-like, in the majority of the cases forming a cluster. Aggregates were manually separated and the thicker individuals fished.

AtPRK crystals showed a bipiramidal morphology and grew in three to five weeks from a reservoir solution containing 1.4 – 1.7 M sodium malonate, pH 5.0. The best diffracting crystal of *AtPRK* was obtained in 1.5 M sodium malonate, pH 5.0.

Crystals were mounted from the crystallization drop into cryo-loops, briefly soaked in a cryo-protectant solution containing 30% w/v PEG MME and 20% v/v PEG 200 for *CrPRK* and 1.7 M sodium malonate, pH 5.0, and 30% v/v glycerol for *AtPRK*, then frozen in liquid nitrogen. Diffraction images were recorded at 100 K at the Elettra synchrotron radiation source (Trieste, beam line XRD1) for *CrPRK* and at the European Synchrotron Radiation Facility (Grenoble, beam line ID14-4) for *AtPRK*. Data collection parameters are reported in Table S4.

The data at a resolution of 2.6 Å for *CrPRK* and 2.5 Å for *AtPRK* were processed using XDS (1) and scaled with SCALA (2). The correct space group was determined with POINTLESS (2) and confirmed in the structure solution stage. Data collection statistics are reported in Table S5.

Structure Solution and Refinement.

CrPRK structure was solved by molecular replacement using the program PHASER (3) from PHENIX (4) starting from the coordinates of PRK from *Methanospirillum hungatei* (PDB code 5B3F) (5) deprived of sulfate ions and water molecules. The protein chain was traced by Autobuilt from PHENIX (4) and Buccaneer (6) from CCP4 package. The refinement was performed with REFMAC 5.8.0135 (7) selecting 5% of reflections for R_{free} . The manual rebuilding was performed with Coot (8). The residual electron density map showed the position of a sulfate ion for each monomer coming from the crystallization solution, which was added to the model. Water molecules were automatically added and, after a visual inspection, confirmed in the model if the relative electron density value in the $(2F_o - F_c)$ maps exceeded $0.19 \text{ e}^{-\text{Å}^{-3}}$ (1.0σ) and if they fell into an appropriate hydrogen bonding environment. The last refinement cycle was performed with PHENIX (4). The final model of *CrPRK* lacks six residues at the C-terminal end of both chains.

The structure of *CrPRK* without sulfate ions and waters, was used as initial model to solve the *AtPRK* structure by molecular replacement using MOLREP (9). The refinement was performed as described for *CrPRK*. The final model of *AtPRK* lacks three residues at the N-terminal domain and seven and ten residues at the C-terminal domain of chain A and B, respectively.

The refinement statistics of both structures are reported in Table S5. All structure figures were prepared using PyMOL (The PyMOL Molecular Graphics System, Schrödinger, LLC).

Small Angle X-ray Scattering Data Collection.

Data collection parameters are reported in Table S4. A Size Exclusion Chromatography SEC-SAXS experiment was performed using a HPLC system (Shimadzu) directly connected to the measurement capillary. A volume of 100 μl of reduced *CrPRK* (6.1 mg ml^{-1}) was loaded onto a Superdex 200 10/300 GL column (GE Healthcare) pre-equilibrated in 50 mM Tris-HCl, 150 mM

KCl, pH 7.5. The sample was eluted at a flow rate of 0.5 ml min^{-1} and SAXS frames obtained by 1 s exposure were collected continuously. The automatic pipeline for SEC-SAXS data analysis implemented at BM29 was used to assess the quality of the collected data (10).

Small Angle X-ray Scattering Data Analysis.

Afterwards, a classification of the collected frames as buffer (0-14 ml) or protein frames (14-17.75 ml) was performed on the basis of the SAXS intensity trace (Fig. S13). Statistical test implemented in CorrMap (11) aided by visual inspection, was used to choose the superimposable buffer intensity profiles. The averaging of the buffer profiles, the subtraction of the averaged buffer intensity from the protein data and an automatic analysis of the subtracted protein profiles was performed with a Matlab script. The script used the tools of the ATSAS package (12) to automatically obtain from the subtracted intensity $I(q)$: (i) the $I(0)$ and the gyration radius (R_g) via the Guinier approximation (13) $I(q) = I(0) \cdot \exp[-(qR_g)^2/3]$; (ii) the pair-distance $[p(r)]$ function, from which the maximum particle dimension (D_{\max}) was estimated, in addition to an independent calculation of $I(0)$ and R_g . Estimates of the MW were also determined both from the Porod invariant (14) as 0.6 times the Porod volume (V_p) for roughly globular particles (12) and by the invariant volume-of-correlation length (V_c), through a power-law relationship between V_c , R_g and MW that has been parametrized (15). The protein frames giving constant R_g values were scaled to the intensity of the elution maximum and averaged in order to obtain a single representative scattering profile with good signal to noise ratio, presented in the results and used for modelling.

Modeling from SAXS Data

The sequence and the homodimeric state of CrPRK were given as inputs in GASBOR and a 2-fold symmetry was imposed in the calculations. A series of 10 models was generated. The similarity of the structures obtained by repeated calculations was checked by DAMAVER (16) in which the superposition is performed by the SUPCOMB code (17).

All programs used for SAXS data analysis and reconstruction, belong to the ATSAS package 2.7 (12). The graphical representations of the obtained three-dimensional models were built by using

PyMOL (The PyMOL Molecular Graphics System, Schrödinger, LLC).

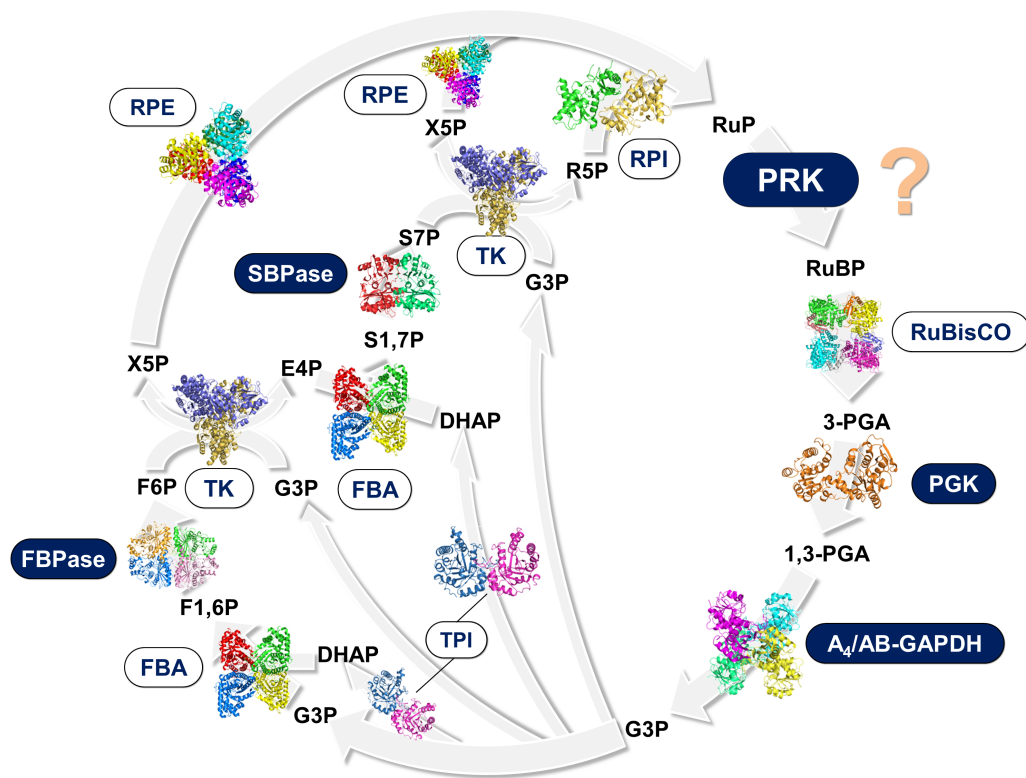


Fig. S1. Schematic representation of the C₄ cycle. The crystal structure of six chloroplast enzymes from different organisms: RuBisCO, ribulose-1,5-bisphosphate Carboxylase/Oxygenase from *Spinacia oleracea* (PDB ID code 1AUS) (18); GAPDH, glyceraldehyde-3-phosphatedehydrogenase from *Arabidopsis thaliana* (PDB ID code 3K2B) (19); TPI, triose phosphate isomerase from *Chlamydomonas reinhardtii* (PDB ID code 4MKN) (20); FBPase, fructose-1,6-bisphosphatase from *Pisum sativum* (PDB ID code 1DCU) (21); TK, transketolase from *Chlamydomonas reinhardtii* (PDB ID code 5ND5) (22); SBPase, sedoheptulose-1,7-bisphosphatase from *Physcomitrella patens* (PDB ID code 5IZ3) (23); RPE, ribulose-5-phosphate 3-epimerase from *Solanum tuberosa* (PDB ID code 1RPX) (24), is shown. For the remaining three enzymes, the crystal structure of the most homologous enzymes from non-photosynthetic organisms is reported: PGK, phosphoglycerate kinase from *Bacillus stearothermophilus* (PDB ID code 1PHP) (25) approximately 60% homologous to *Chlamydomonas reinhardtii* PGK1 (sequence accession number A8JC04); FBA, fructose-1,6-bisphosphatealdolase from *Toxoplasma gondii* (PDB ID code

5TJS) (26) approximately 55% homologous to *Arabidopsis thaliana* FBA1 (sequence accession number Q9SJU4); RPI, ribose-5-phosphate isomerase from *Toxoplasma gondii* (PDB ID code 4NML) approximately 50% homologous to *Spinacia oleracea* RPI (sequence accession number Q8RU73).

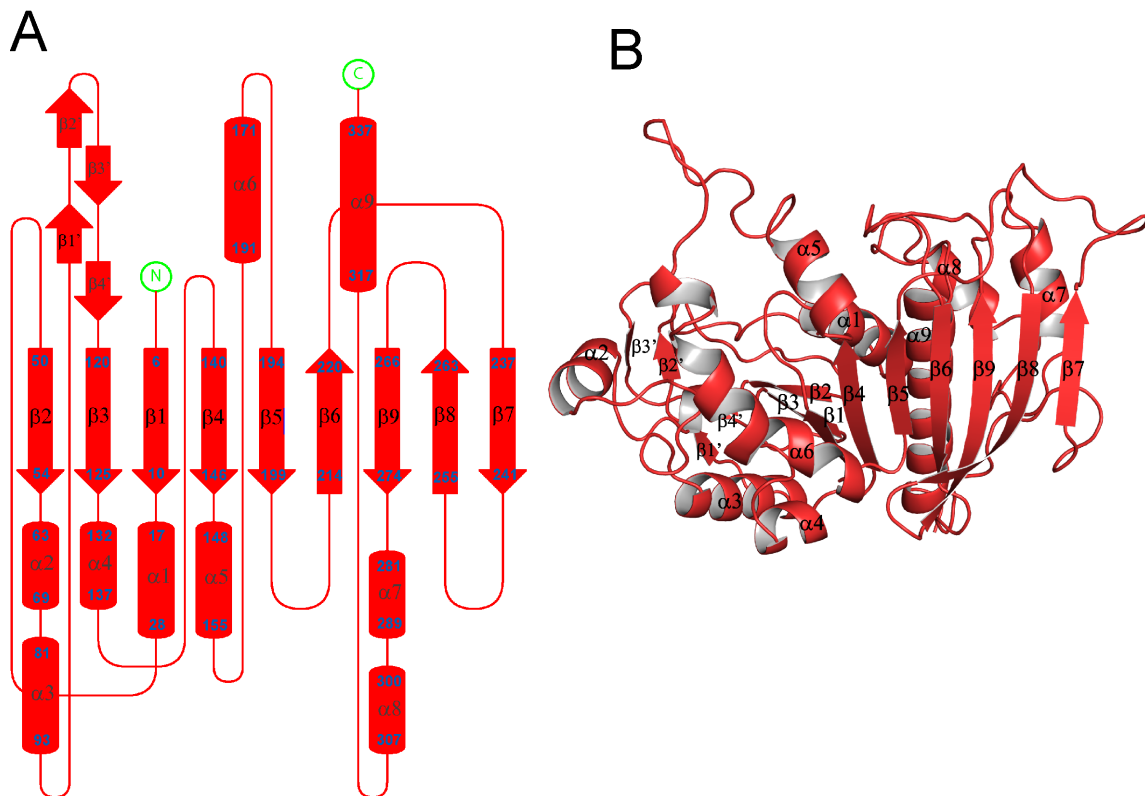


Fig. S2. Crystal structure of *AtPRK*'s monomer. (A) Topology diagram of *AtPRK*. Similarly to *CrPRK* (Fig. 2A), the monomer is composed by a mixed β -sheet of nine strands, by nine α -helices and four additional small β -strands indicated by β' . (B) Cartoon representation of the monomer structure of *AtPRK*. Similarly to *CrPRK* (Fig. 2B) the central β -sheet is sandwiched between helices $\alpha 3$, $\alpha 4$ and $\alpha 6$ and helices $\alpha 1$, $\alpha 7$, $\alpha 8$ and $\alpha 9$. The right end of the monomer consists of stand $\beta 7$ involved in the dimer interface, while the four additional β -strands ($\beta 1'$ to $\beta 4'$) form the left external end of the dimer.

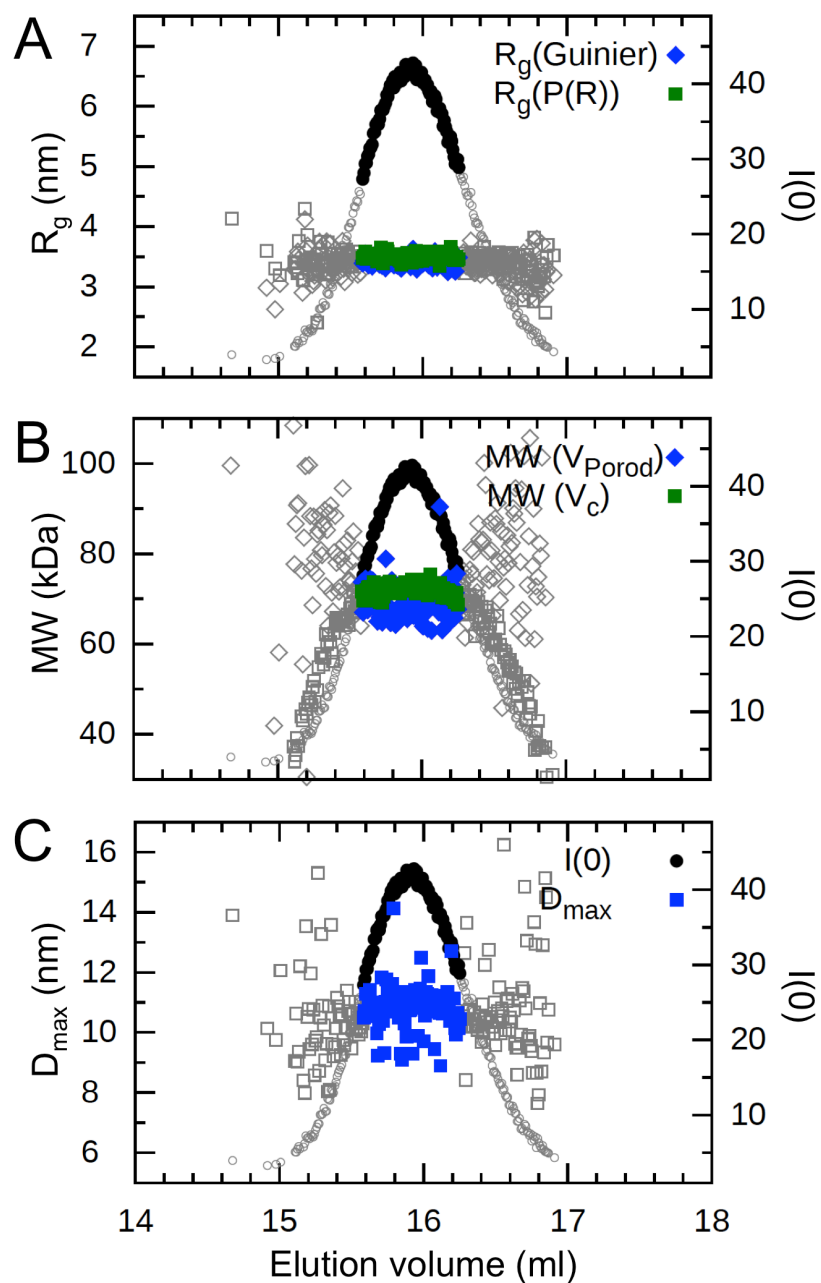


Fig. S3. Parameters determined by SEC-SAXS analysis of CrPRK. (A) $I(0)$ trace (dots) and R_g determined by the Guinier approximation (diamonds) and R_g calculated from the $P(r)$ function (squares). (B) $I(0)$ trace (dots) and MW estimated from the Porod volume (diamonds) and from the volume-of-correlation (squares). (C) $I(0)$ trace (dots) and D_{max} estimated from the $P(r)$ function (squares). The frames used in the average to obtain the representative scattering profile, are highlighted in black compared to the grey neglected frames.

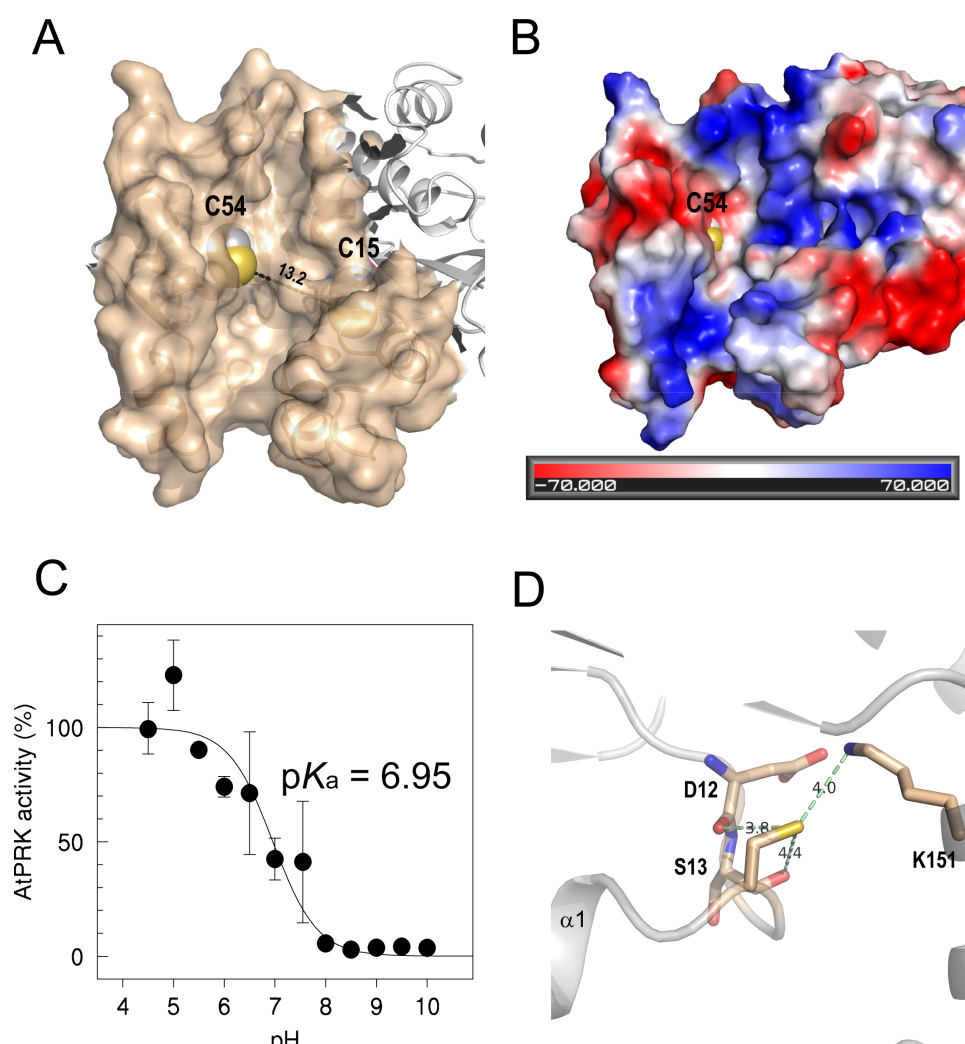


Fig. S4. Active site and TRX-dependent regulation of AtPRK. (A) The catalytic cavity is shown. The distance between the regulatory cysteines is higher than 13 Å. (B) Catalytic cavity electrostatic surface potential. The bottom of the catalytic cavity is marked by a positive potential. The negative potential region observed on the left side of the cavity, is suggested to be involved in the correct positioning of TRX close to regulatory cysteines. (C) The pK_a of Cys15 was determined by measuring the IAM-mediated inactivation as a function of pH. (D) Molecular environment of Cys15 considering a sphere of 5 Å centered on its thiol group. The hydrogen bonds between the thiol group and the neighboring residues and the corresponding distances, are shown.

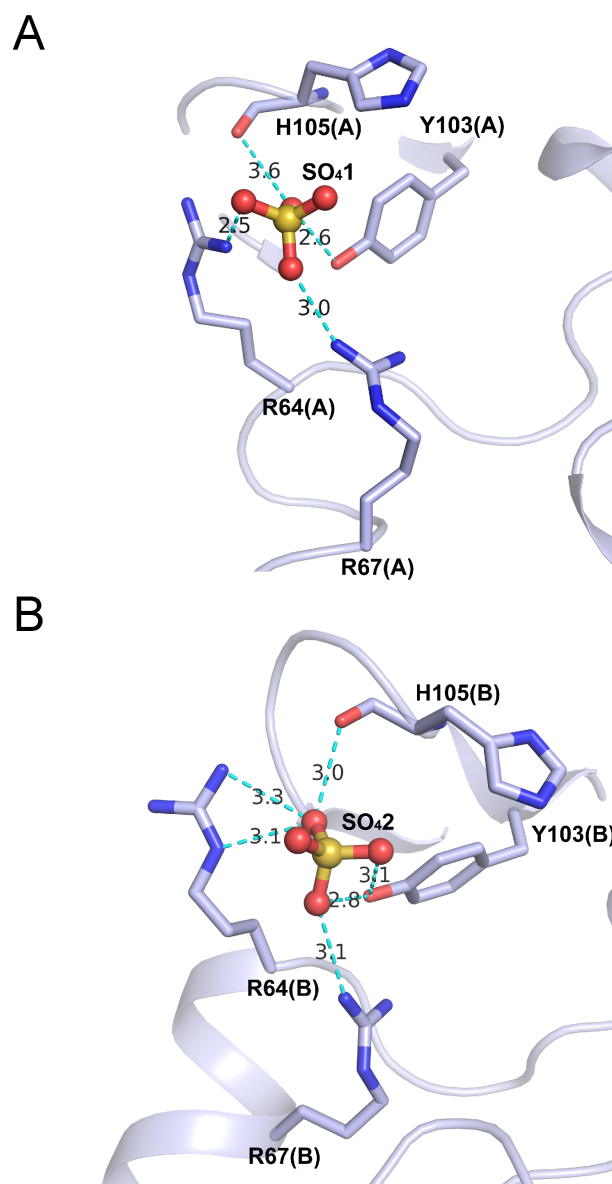
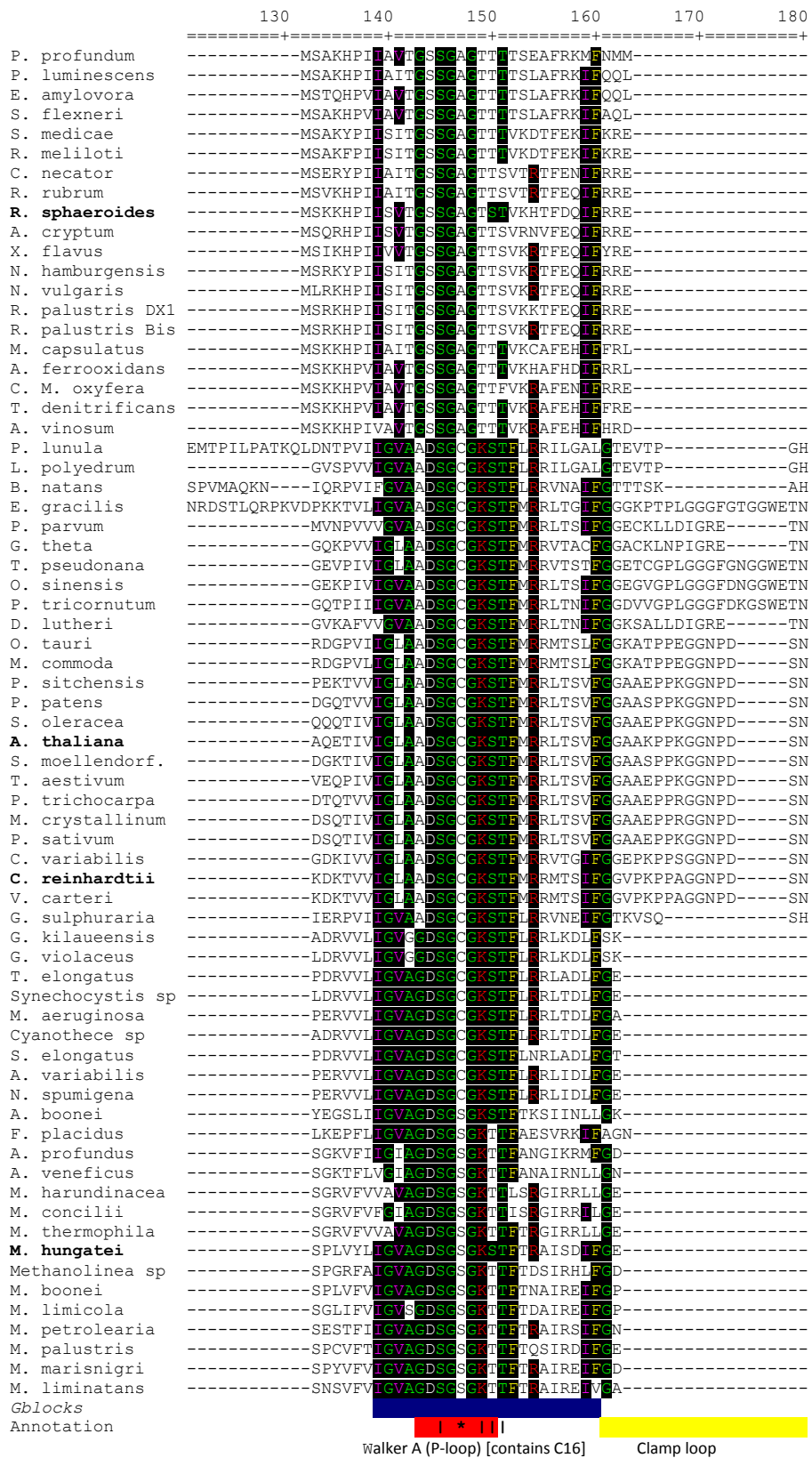


Fig. S5. Binding site of the sulfate ion in *CrPRK*. The interactions between a sulfate ion coming from the crystallization solution, and protein residues in (A) subunit A and (B) subunit B, are represented. Salt-bridges are formed with Arg64 and Arg67, and hydrogen bonds with Tyr103 and His105.

A

| | 10 | 20 | 30 | 40 | 50 | 60 |
|-----------------------|-------------------------------------------------------------|----|----|----|----|-------------------------------------------------------|
| P. profundum | | | | | | |
| P. luminescens | | | | | | |
| E. amylovora | | | | | | |
| S. flexneri | | | | | | |
| S. medicae | | | | | | |
| R. meliloti | | | | | | |
| C. necator | | | | | | |
| R. rubrum | | | | | | |
| R. sphaeroides | | | | | | |
| A. cryptum | | | | | | |
| X. flavus | | | | | | |
| N. hamburgensis | | | | | | |
| N. vulgaris | | | | | | |
| R. palustris DX1 | | | | | | |
| R. palustris Bis | | | | | | |
| M. capsulatus | | | | | | |
| A. ferrooxidans | | | | | | |
| C. M. oxyfera | | | | | | |
| T. denitrificans | | | | | | |
| A. vinosum | | | | | | |
| P. lunula | | | | | | MARSRSALFPALAAALLLWAREAFL |
| L. polyedrum | | | | | | MARSRSVVLPLAAAVALVLLGHVAVFPAPRP |
| B. natans | | | | | | KTNVYAIASVVANVALVGVLLYTQTGEVGSVAASSRVHVAPRMVAPKFSMSGL |
| E. gracilis | | | | | | MMRYDQLIHEGPRQSSPSLLQLASVSATVFCVAFG |
| P. parvum | | | | | | MLSLSVGLSVAF |
| G. theta | | | | | | MGTPLASRSSFASPVAF |
| T. pseudonana | | | | | | |
| O. sinensis | | | | | | MKLSLGIIVALMAASASAFAPTAFA |
| P. tricornutum | | | | | | MKFAVFAFLTATAAFAPTAFA |
| D. lutheri | | | | | | MRRSRVATPVQR |
| O. tauri | | | | | | |
| M. commoda | | | | | | MASMTMSAACAQPLAA |
| P. sitchensis | | | | | | MACPAPTATSTPATVRSWNSQFCSNSP |
| P. patens | | | | | | MAAMACTSAASTALSTPRLAAV |
| S. oleracea | | | | | | MAVCTVYTIPTTHLGSSEFNQN |
| A. thaliana | | | | | | MAVSTIYSTQALNSTHFLTSSS |
| S. moellendorff. | | | | | | MAAACSSSGSSSRIGWFQASSS |
| T. aestivum | | | | | | MAFCSPHTTTSLRSECTT |
| P. trichocarpa | | | | | | MAVCTVYTTQSLNSTCISITPT |
| M. crystallinum | | | | | | MAVSAYTVPT |
| P. sativum | | | | | | |
| C. variabilis | | | | | | MQTAAAVQCKPVCRFMAARPHVARRPAQLAASA |
| C. reinhardtii | | | | | | |
| V. carteri | | | | | | MAFTMRAPAPRAT |
| G. sulphuraria | MELRDSLSTMGFVLPSTTSYVVYKSSSTIKSKKYNKRNYSVSRVQVGGSKISSWSSNRL | | | | | |
| G. kilauensis | | | | | | |
| G. violaceus | | | | | | |
| T. elongatus | | | | | | |
| Synechocystis sp | | | | | | |
| M. aeruginosa | | | | | | |
| Cyanothece sp | | | | | | |
| S. elongatus | | | | | | |
| A. variabilis | | | | | | |
| N. spumigena | | | | | | |
| A. boonei | | | | | | |
| F. placidus | | | | | | |
| A. profundus | | | | | | |
| A. veneficus | | | | | | |
| M. harundinacea | | | | | | |
| M. concilii | | | | | | |
| M. thermophila | | | | | | |
| M. hungatei | | | | | | |
| Methanolinea sp | | | | | | |
| M. boonei | | | | | | |
| M. limicola | | | | | | |
| M. petrolearia | | | | | | |
| M. palustris | | | | | | |
| M. marisnigri | | | | | | |
| M. liminatans | | | | | | |
| Gblocks | | | | | | |
| Annotation | | | | | | |

| | 70 | 80 | 90 | 100 | 110 | 120 |
|-----------------------|---------------------------------------------------------------|-------|-------|-------|-------|-------|
| P. profundum | ----- | ----- | ----- | ----- | ----- | ----- |
| P. luminescens | ----- | ----- | ----- | ----- | ----- | ----- |
| E. amylovora | ----- | ----- | ----- | ----- | ----- | ----- |
| S. flexneri | ----- | ----- | ----- | ----- | ----- | ----- |
| S. medicae | ----- | ----- | ----- | ----- | ----- | ----- |
| R. meliloti | ----- | ----- | ----- | ----- | ----- | ----- |
| C. necator | ----- | ----- | ----- | ----- | ----- | ----- |
| R. rubrum | ----- | ----- | ----- | ----- | ----- | ----- |
| R. sphaeroides | ----- | ----- | ----- | ----- | ----- | ----- |
| A. cryptum | ----- | ----- | ----- | ----- | ----- | ----- |
| X. flavus | ----- | ----- | ----- | ----- | ----- | ----- |
| N. hamburgensis | ----- | ----- | ----- | ----- | ----- | ----- |
| N. vulgaris | ----- | ----- | ----- | ----- | ----- | ----- |
| R. palustris DX1 | ----- | ----- | ----- | ----- | ----- | ----- |
| R. palustris Bis | ----- | ----- | ----- | ----- | ----- | ----- |
| M. capsulatus | ----- | ----- | ----- | ----- | ----- | ----- |
| A. ferrooxidans | ----- | ----- | ----- | ----- | ----- | ----- |
| C. M. oxyfera | ----- | ----- | ----- | ----- | ----- | ----- |
| T. denitrificans | ----- | ----- | ----- | ----- | ----- | ----- |
| A. vinosum | ----- | ----- | ----- | ----- | ----- | ----- |
| P. lunula | LPRSSPQAPRASSARVPKVVALRAAADSQPTGLVWFSPEAEAKLREVDGIKLYPTHAWTE | | | | | |
| L. polyedrum | RAARALTAMRGAANSVPTGLVWFSPEAEAKLREEDGIKMYPTHAWTDDMMPIVPATKE-- | | | | | |
| B. natans | RPAQREARVAAHGKGVVSTVALDAPVATSEDSMTWSSAKGNEVDAGMGQVHIEAAISNEV | | | | | |
| E. gracilis | IGYTFSSGGLVENYATTRPVQTQPAALILPKAVRYAGVSGQPQVESREARTALHAAATGTV | | | | | |
| P. parvum | SPA-----PSVVSQSRVAPRATVVE-- | | | | | |
| G. theta | -----RVTNDKEMDVAIQQISMSLAS-- | | | | | |
| T. pseudonana | -----MKFLVASLIASAFSTINTPTLRGNSALAALKD-- | | | | | |
| O. sinensis | -----MPANTLRAAAPASPSALNMALKE-- | | | | | |
| P. tricornutum | VPS-----NLRGVAPSASSLNMALKE-- | | | | | |
| D. lutheri | VPQ-----TACTVLMATKT-- | | | | | |
| O. tauri | -----MSASLGFSTSVRAAPVRATRDVAKPRATRARVTTAK-- | | | | | |
| M. commoda | RKFQGS-----RVSGKSVVRAAKRNVTVKAE-- | | | | | |
| P. sitchensis | LAS-----FSLAHTTRRPRRALVVCVSG-- | | | | | |
| P. patens | KPARSARPVH-----LTSAFHGQSVASVSQVAGFESSGVKYSAGRRAVVVCKAA-- | | | | | |
| S. oleracea | NKQVFFNYKR-----SSSNNTLFTTRPSYVITCS-- | | | | | |
| A. thaliana | SSKQVF-----LYRRQPQTNRFRNTLITC-- | | | | | |
| S. moellendorf. | SSNIGILQHHPWRGSHASSLFSFPKLGARIGSSSGSGSSGTSSSNRVRVLVCCAAGG-- | | | | | |
| T. aestivum | IPNSGFRQNG-----VIFFTTRSSRSNTRHGARTFQVSCA-- | | | | | |
| P. trichocarpa | KTHLGFNQRH-----VVFYSTNKKTTKRASSAVITCSA-- | | | | | |
| M. crystallinum | TSHLGFNQKK-----QLFFCNKSAYKRVSFSSRPCVITCLAG-- | | | | | |
| P. sativum | -----AG-- | | | | | |
| C. variabilis | APIARTFSS-----TLNQRTLKAGRVASRVVVVKAEG-- | | | | | |
| C. reinhardtii | -----MAFTMRAPAPRATAQSRVTANRARRSLVVRAD-- | | | | | |
| V. carteri | -----AQSRVTASRASRRVLVVKAQ-- | | | | | |
| G. sulphuraria | LFQGKEINTTRKTTTKYWIITVSSQTALEEFVNCSSGAKGVHESAGISRSKSKVLNRNG-- | | | | | |
| G. kilaeuensis | -----MVSK-- | | | | | |
| G. violaceus | -----MVST-- | | | | | |
| T. elongatus | -----MSSK-- | | | | | |
| Synechocystis sp | -----MTTQ-- | | | | | |
| M. aeruginosa | -----MANK-- | | | | | |
| Cyanothece sp | -----MTTQ-- | | | | | |
| S. elongatus | -----MSK-- | | | | | |
| A. variabilis | -----MTTK-- | | | | | |
| N. spumigena | -----MTTK-- | | | | | |
| A. boonei | -----MLGEFRRRLEE-- | | | | | |
| F. placidus | -----MILEK-- | | | | | |
| A. profundus | -----MLKEKLIK-- | | | | | |
| A. veneficus | -----MTSNLKERLKE-- | | | | | |
| M. harundinacea | -----MAERLKG-- | | | | | |
| M. concilii | -----MRSCLKDRIRE-- | | | | | |
| M. thermophila | -----MRLLEKIRE-- | | | | | |
| M. hungatei | -----MSQPENFREVIRH-- | | | | | |
| Methanolinea sp | -----MTSKPGFKEI IKS-- | | | | | |
| M. boonei | -----MPRTPPFKEI IAR-- | | | | | |
| M. limicola | -----MSCIDEYLSESGKCRNRFSGDDEYCFSSSGKGINLKKI IEN-- | | | | | |
| M. petrolearia | -----MDYSHETNLKNFRHAVDS-- | | | | | |
| M. palustris | -----MMQTEGKTGEKPDLCPGTGGLNFKDRIAS-- | | | | | |
| M. marisnigri | -----MPPSDFKRVIAE-- | | | | | |
| M. liminatans | -----MDTPVFRDLISG-- | | | | | |
| Gblocks | | | | | | |
| Annotation | | | | | | |



| | 190 | 200 | 210 | 220 | 230 | 240 |
|-----------------------|----------------------|------------------------|------------|-------------------|-----|-----|
| P. profundum | ---NINASWLEGDSEFRYTP | PEMDVEIRKAKEQGR-HISYFG | PELND | FPQLEKFFRQYG | | |
| P. luminescens | ---DISAAQIEGDSEFRYTP | PEMDAAIRKAKEQGR-HISYFG | PELND | FGMLEKTMIDYG | | |
| E. amylovora | ---GLHAAEIEGDSEFRFT | PEMDAIRKARDMGK-HVSYFG | PELND | FALERTFSEYG | | |
| S. flexneri | ---NLHAAEVEGDSEFRYTP | PEMDAIRKARDAGR-HISYFG | PELND | FGLEQTFIEYG | | |
| S. medicae | ---NISASFIEGDSEFRYD | RETMRSKIAEEKARGV-DF | HFSAE | NELEILESVPFAEYG | | |
| R. meliloti | ---NISASFIEGDSEFRD | RETMRSKIAEEKARGV-DF | HFSAE | NELEILESVPFAEYG | | |
| C. necator | ---GVKSVVIEGDSEFRYD | RAEMVVKMAEAERTGNMNF | SHFGEEN | NLGELENLFRSYA | | |
| R. rubrum | ---GVNAAVVEGDSEFRN | DKAMVIAMAEAQKAGNANF | SHFG | PELNFEELETLFRTYG | | |
| R. sphaeroides | ---GVKAVSIEGDSEFRN | ADMVAELDRRYAAGDATF | SHFSYE | ANELKELERVFREYG | | |
| A. cryptum | ---KITAAHIEGDSEFRYD | RAEMTKMAEAEAAGNRHFS | SHFS | PETNLLAELAATFESYA | | |
| X. flavus | ---KVKAAFVEGDSEFRYD | YEMRELMAEAAKGNKHF | SHFS | PETNRLDLAQLFKDYG | | |
| N. hamburgensis | ---NVVAAYIEGDSEFRYN | ADMRTMAEESDRGNKHF | SHFS | PETNLFELEAVFRSYS | | |
| N. vulgaris | ---NVVAAYIEGDSEFRYN | ADMRTMAEESDKGNKHF | SHFS | PETNLFELEAGVFRSYG | | |
| R. palustris DX1 | ---NVNAAYIEGDSEFRYN | VDMRNKMAEAEAERGNR | SHFS | PETNLFELEQTFKSYA | | |
| R. palustris Bis | ---NVNAAYIEGDSEFRYN | VDMRTMAEAEAEGNKT | SHFS | PETNLFELETTFRDYS | | |
| M. capsulatus | ---GLKPLVIEGDSEFRYD | VEMRAQIDKARREG-RHF | SHFSIE | NILFELENVFRHYG | | |
| A. ferrooxidans | ---KIDFVIEGDSEFRYN | NEMREAIKAAADG-KTIS | SHFG | EGNDFEALERLFRFYG | | |
| C. M. oxyfera | ---EITSAVIEGDSEFRV | TAQFKERSAVEH----NF | SHFG | PELNFHALEALFKSYG | | |
| T. denitrificans | ---KINAAVIEGDSEFRSL | AVFHEAVKKAEEAGNF | SHFG | PELNFHKLALFKTYG | | |
| A. vinosum | ---NISAAVIEGDSEFRS | YATMAEMAAPAKRGE-SL | SHFG | PELNFHLELFRFYG | | |
| P. lunula | TAVGDMMLVICLDD-YH | TNDRAGRKA-----T | LHADAREND | FALMGSQIEAL | | |
| L. polyedrum | TAIGDMMVICLDD-YH | TNDRAGRKA-----T | LHADAKEND | FALMGVQIEAL | | |
| B. natans | TLPTGDLLVICLDD-FH | TLDRTGAD-----T | LISADVRR | ANFALMAQDLK | | |
| E. gracilis | TLVSDKTVVICLDD-YH | LNDRAGRKA-----T | LHADQREND | FALMFSQMSK | | |
| P. parvum | TLVSDMTVICLDD-YH | KWDTGRKSNPEWPN--- | LHAEACQDW | KMAADVLDL | | |
| G. theta | TLISDMTVICLDD-YH | LNDRGRK-----T | LHADREND | FALMYSQVKAL | | |
| T. pseudonana | TLVSDMAVICLDD-YH | LNDREGRKA-----S | LHANTAEOK | FALMHVKAL | | |
| O. sinensis | SLVSDLTVICLDD-YH | LNDRGRK-----T | QRHADREND | FALMYSQIAAL | | |
| P. tricornutum | TLVSDLTVICLDD-YH | LNDRGRK-----T | MRHADREND | FALMYSQVKAL | | |
| D. lutheri | TLVSDKTVVICLDD-YH | LYDRKGS-----N | KLHAKCQK | WDLMAQVAAT | | |
| O. tauri | TLISETTVICLDD-YH | LNDRGRK-----S | LHNLKEQN | FALMYSQVKAL | | |
| M. commoda | TLISDSTVICLDD-YH | LNDRGRK-----S | LHNLKEQN | FALMYSQTKAL | | |
| P. sitchensis | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| P. patens | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| S. oleracea | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| A. thaliana | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| S. moellendorf. | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| T. aestivum | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| P. trichocarpa | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| M. crystallinum | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| P. sativum | TLISDTTVICLDD-YH | SLDRYGRK-----K | VHADDRAND | FALMYSQVKAL | | |
| C. variabilis | TLISDMTVICLDD-YH | SLDRYGRK-----A | VHADDRAND | FALMYSQVKAL | | |
| C. reinhardtii | TLISDMTVICLDD-YH | CLDRNGRKA-----K | VHADDRAND | FALMYSQVKAL | | |
| V. carteri | TLISDMTVICLDD-YH | CLDRNGRKA-----K | VHADDRAND | FALMYSQVKAL | | |
| G. sulphuraria | TPQGELVVICLDD-FH | TLDRKGRKAE-----K | VHADDRAND | FALMYSQIAAL | | |
| G. kilaeuensis | ---ELVVICLDD-YH | SLDRKGRK-----T | LHADDRAND | FALMYSQAAAL | | |
| G. violaceus | ---ELVVICLDD-YH | SLDRKGRK-----T | LHADDRAND | FALMYSQATAAL | | |
| T. elongatus | ---DFMVICLDD-YH | SLDRKGRK-----M | LHADDRAND | FALMYSQIKAL | | |
| Synechocystis sp | ---EFMVICLDD-YH | SLDRQGRKA-----A | VHADDRAND | FALMYSQIKT | | |
| M. aeruginosa | ---EFMVICLDD-YH | CLDRKGRK-----V | VHADDRAND | FALMYSQIKAL | | |
| Cyanotheca sp | ---EFMVICLDD-YH | SLDRQGRK-----A | VHADDRAND | FALMYSQIKAL | | |
| S. elongatus | ---ELMVICLDD-YH | SLDRKGRK-----A | VHADDRAND | FALMYSQVKAL | | |
| A. variabilis | ---EFMVICLDD-YH | SLDRKGRK-----T | LHADDRAND | FALMYSQIKAL | | |
| N. spumigena | ---EFMVICLDD-YH | SLDRKGRK-----T | LHADDRAND | FALMYSQIKAL | | |
| A. boonei | ---DLVSSFTLDD-YH | TEDETNRK-----T | LHLPDRKTN | NLKLAAEHL | | |
| F. placidus | ---VATITLDD-YH | KYGRKGRK-----L | LHPLNPEMTN | LDLFRHLKLIK | | |
| A. profundus | ---DIVSHITLDD-YH | VYDREMER-----L | LHPLHESANN | KLVEETHLYL | | |
| A. venificus | ---DIVSITLDD-YH | VYDQTEWK-----L | LHPLHESANN | KLVEETHLML | | |
| M. harundinacea | ---EAVSTFSMDD-YH | SLDREERV-----R | LHPLNPEANR | LDLALHLRSLR | | |
| M. concilii | ---DMVATFSMDD-YH | SLDRGRK-----R | LHPLHESANN | QFDLALHLEAR | | |
| M. thumaphila | ---DVSSTFSMDD-YH | SLDRGRK-----L | LHPLHESANN | RFDLALHLMAR | | |
| M. hungatei | ---ELVSSITVDD-YH | LYDRKRTSE-----M | LHPLHESANN | KLLENLMDL | | |
| Methanolinea sp | ---SMVATITLDD-YH | IYNTQDE-----I | LHPLHESANN | LAGLERDLRL | | |
| M. boonei | ---DLVATITLDD-YH | TYDREHH-----L | LHPLHESANN | LSGLEADVHL | | |
| M. limicola | ---SFVSTITLDD-YH | ILDREH-----K | LHPLHESANN | LSLEHLSDL | | |
| M. petrolearia | ---NMVITLSDDD-YH | VLDREKR-----T | LHPLHESANN | LSLEHVAL | | |
| M. palustris | ---DLVITLSDDD-YH | LYDREK-----R | LHPLHESANN | RLDLEHDLVLT | | |
| M. marisnigri | ---DLVSTITLDD-YH | RYDRQK-----L | LHPLHESANN | RFDLALHLEAR | | |
| M. liminatans | ---DLVATITLDD-YH | RYDRK-----L | LHPLHESANN | LDLADHIRAL | | |

Annotation * + # #

C55

| | 250 | 260 | 270 | 280 | 290 | 300 |
|-----------------------|-----------|----------------------------|-------|---------------|-----------------|--------------|
| P. profundum | DDGSGQFR | YLFDEAVP | ----- | YNQMP | SFTFWQKLPENTDV | FYFGLGGV |
| P. luminescens | ETGEGRRR | YLYDDAVP | ----- | YNQLP | TFTFWESLPKQTDV | FYFGLGGV |
| E. amylovora | RSGKGKSR | YLYDEAVP | ----- | WNQVP | TFTFWQPLAE | TDVIFYEGLGGV |
| S. flexneri | QSGKGKSR | YLYDEAVP | ----- | WNQVP | TFTFWQPLPE | TDVIFYEGLGGV |
| S. medicae | RRGVGRTR | YVDDAEAVK | ----- | FGSDP | TFTDWEFR-DSDL | FYFGLGCA |
| R. meliloti | RRGVGRTR | YVDEAEAAK | ----- | FGSDP | TFTDWEFR-DSDL | FYFGLGCA |
| C. necator | ETGTGMHR | YLSPEEAAP | ----- | FGQEP | TFTQWELPADTDL | FYFGLGGV |
| R. rubrum | ETGGRRRL | YLNDEEAAP | ----- | FAQEP | TFTFWEDLP-ESDL | FYFGLGAV |
| R. sphaeroides | ETGQRTRT | YVDDAEAAAR | ----- | TGVAP | NFTDWRDFSDSHL | FYFGLGAV |
| A. cryptum | ATGTGQYR | YIYDQDEAER | ----- | YGGTP | TFTFWEDLPA | TDILLYEGLGAV |
| X. flavus | ATGSGRFR | YVDDAGEAKL | ----- | YNTEP | RFTDWELEQGTDL | FYFGLGAV |
| N. hamburgensis | ESGTGNTRY | YVDDVESAK | ----- | HGVPP | TFTDWQALPENSDL | FYFGLGAV |
| N. vulgaris | ETGTGNTRY | YVDDAESAL | ----- | HGVPP | TFTDWQPLPASDL | FYFGLGAV |
| R. palustris DX1 | ETGTGRTR | YVDDDEEAAL | ----- | HGVPP | NFTGWRDLDPQSDL | FYFGLGAV |
| R. palustris Bis | ETGTGKTR | YVDDKEAAI | ----- | HGVQP | NFTDWQTLPEGSDL | FYFGLGAV |
| M. capsulatus | ETGTARRR | YVNEAESQRL | ----- | GGYKP | TFTFWEEVPAAGTDL | FYFGLGGV |
| A. ferrooxidans | ESGHQMR | YVDELEAEL | ----- | RGSAP | TFTFWQDIPLGTDL | FYFGLGGV |
| C. M. oxyfera | EAGVGKRY | YLNNAKEAAFHCKRLADAGVTCNADS | ----- | EFTFWEDIESNTD | ILIFYEGLGLV | |
| T. denitrificans | ETGGGKRY | YISDEEADQHNKRLN | ----- | TSLNP | EFTFWEDVPPGSDV | FYFGLGLV |
| A. vinosum | ETGCGKRY | YISDEEARQLNARLG | ----- | TSLNP | EFTFWEDLPAGTDL | FYFGLGLV |
| P. lunula | QCKAVY | KPFLYNDT | ----- | CFKDPPELIE | FNKVMVF | EGLHFIY |
| L. polyedrum | QCKAVY | KPFLYNDT | ----- | CFKDPPELIE | FNKVMVF | EGLHFIY |
| B. natans | QCRRAIK | KPFLYNDT | ----- | CAIDPPEVETIH | FNHIIIVE | EGLHML |
| E. gracilis | RCEETIA | KPFLYNVN | ----- | CTLDPEEIA | FASMIIE | EGLHLL |
| P. parvum | AQKSVS | KPFLYNVV | ----- | GELDPYEDVD | FTPVIVIE | EGLHMY |
| G. theta | ECKKVM | KPFLYNVN | ----- | CTLDEAEIT | FTPVIVIE | EGLHMY |
| T. pseudonana | ECKTIM | KPFLYNVN | ----- | CTLDPEEIE | FTPVIVIE | EGLHMY |
| O. sinensis | NCESTIE | KPFLYNVN | ----- | CTLDPEEIV | FTPVIVIE | EGLHMY |
| P. tricornutum | DCKTVE | KPFLYNVN | ----- | CTLDPEEIE | FTPVIVIE | EGLHMY |
| D. lutheri | AANSVM | KPFLYNVN | ----- | GELDPEEIV | FTPVIVIE | EGLHML |
| O. tauri | ECKSVD | KPFLYNVV | ----- | GVFDPAEKIE | SPEVIVIE | EGLHFA |
| M. commoda | ECKAVD | KPFLYNVV | ----- | GVFDPAEKIE | SPSIVIE | EGLHFA |
| P. sitchensis | ECKSDM | KPFLYNVV | ----- | GILDPAEQIN | FPKIVIVIE | EGLHMF |
| P. patens | ECKSVE | KPFLYNVV | ----- | GLLDPAPEITH | FPKIVIVIE | EGLHMF |
| S. oleracea | ECKAVD | KPFLYNVV | ----- | GLLDPPPELIQ | FPKIVIVIE | EGLHMY |
| A. thaliana | NGLIAVE | KPFLYNVV | ----- | GLLDPPPELIQ | FPKIVIVIE | EGLHMF |
| S. moellendorf. | ECKAVQ | KPFLYNVV | ----- | GLLDPPPELIQ | FPKIVIVIE | EGLHMF |
| T. aestivum | ECKAIE | KPFLYNVV | ----- | GLLDPAPELIQ | FPKIVIVIE | EGLHMY |
| P. trichocarpa | DCTAVE | KPFLYNVV | ----- | GLLDPPPELIK | FPKIVIVIE | EGLHMY |
| M. crystallinum | ECKAVE | KPFLYNVV | ----- | GLLDPAPELIK | FPKIVIVIE | EGLHMF |
| P. sativum | DCKSVQ | KPFLYNVV | ----- | GLLDPAPELIK | FPKIVIVIE | EGLHMY |
| C. variabilis | ECKAVD | KPFLYNVV | ----- | GLLDPPPEPTS | SPNIVIVIE | EGLHMY |
| C. reinhardtii | ECKSVD | KPFLYNVV | ----- | GLLDPAPEKIE | SPPIVIVIE | EGLHMY |
| V. carteri | ECKAVD | KPFLYNVV | ----- | GLLDPAPEKID | SPNIVIVIE | EGLHMY |
| G. sulphuraria | EQYDIM | KPFLYNET | ----- | GLIDPPPELIQ | FNHIIIVE | EGLHMY |
| G. kilauensis | SQSIQ | KPFLYNET | ----- | GKIDPPPERID | FTPVIVIE | EGLHMY |
| G. violaceus | VQSIM | KPFLYNET | ----- | GKIDPPPELIE | FTPVIVIE | EGLHMY |
| T. elongatus | NCESIM | KPFLYNET | ----- | CTIDPPPEKVD | FNHIVIVIE | EGLHMY |
| Synechocystis sp | SQSIM | KPFLYNET | ----- | GLLDPPPEKVE | FNKVVIVIE | EGLHMY |
| M. aeruginosa | GGQAIN | KPFLYNET | ----- | GMDPPPEIE | FNKVIVIE | EGLHMY |
| Cyanotheca sp | EQQPIM | KPFLYNET | ----- | GMDPPPERIE | FNKVIVIE | EGLHMY |
| S. elongatus | NCETIM | KPFLYNET | ----- | GLIDPPPEKIE | FNHIIIVE | EGLHMY |
| A. variabilis | EQQTIN | KPFLYNET | ----- | GLIDPPPEIVK | FNHVVIVIE | EGLHMY |
| N. spumigena | EQQVIQ | KPFLYNET | ----- | GMDPPPERVE | FNHIIIVE | EGLHMY |
| A. boonei | KQNAII | KPFLYNET | ----- | GKFDPPPEVFE | FKKIVIVIE | EGLHMY |
| F. placidus | EWKEFE | KPFLYNET | ----- | GEVKCCIFK | BEKVIVIE | EGLHMY |
| A. profundus | KQEKIK | KPFLYNET | ----- | CTFGWEDFE | STPVIVIE | EGLHMY |
| A. veneficus | KQETIR | KPFLYNET | ----- | CTFGWEDFT | STPVIVIE | EGLHMY |
| M. harundinacea | LQETIA | KPFLYNET | ----- | GEFREVPFR | SGPVIVIE | EGLHMY |
| M. concilii | RNERID | KPFLYNET | ----- | GEISGIVPFG | FAPVIVIE | EGLHMY |
| M. thermophila | RGLAIE | KPFLYNET | ----- | GEIRGIVIFK | FSPVIVIE | EGLHMY |
| M. hungatei | AQRTIQ | KPFLYNET | ----- | CTFGEPELFS | BTKEIVIE | EGLHMY |
| Methanolinea sp | QCKGIY | KPFLYNET | ----- | GRLEGPEYLA | FAKIVIVIE | EGLHMY |
| M. boonei | QCYAIE | KPFLYNET | ----- | CTFDPPPIFPF | SKKIVIVIE | EGLHMY |
| M. limicola | SKKEIL | KPFLYNET | ----- | GKFDPEVPFS | SSKIVIVIE | EGLHMY |
| M. petrolearia | ECKSID | KPFLYNET | ----- | SKIEGVPVRLS | FSRIVIVIE | EGLHMY |
| M. palustris | EERTID | KPFLYNET | ----- | GRFAPPIRFT | EKKIVIVIE | EGLHMY |
| M. marisnigri | AQRTIE | KPFLYNET | ----- | GRFDPPVPFS | FTKIVIVIE | EGLHMY |
| M. liminatans | SENTVM | KPFLYNET | ----- | CTFDPPPIPFR | FARVIVIE | EGLHMY |
| <i>Gblocks</i> | | | | | | |
| Annotation | # | # | | | | Walker B |

| | 310 | 320 | 330 | 340 | 350 | 360 |
|-----------------------|--------------|----------------|---------------|--------------|------------|--------------|
| P. profundum | VD----- | GEVNAAEHVDLL | GMVPIVNWLEWIQ | KVVDTRD | RGHREAIMES | IVRSMDD |
| P. luminescens | VT----- | PQHNVAHVLLVGV | PIVNWLEWIQ | LIIRDITGER | RGHQEAIMDS | VVRSMDD |
| E. amylovora | VT----- | PLHNVAENVLLVGV | PIVNWLEWIQ | LVIRDITSE | RGHREAIMDS | VVRSMDD |
| S. flexneri | VT----- | PQHNVAQHVLLVGV | PIVNWLEWIQ | LIIRDITSE | RGHREAIMDS | VVRSMDD |
| S. medicae | VT----- | DTINLAQHCDLKI | GVVPIVNWLEWIQ | KIHRDKAATRGY | TEAATD | TLRSMDD |
| R. meliloti | VT----- | DTVNLAQHCDLKI | GVVPIVNWLEWIQ | KIHRDKAATRGY | TEAATD | TLRSMDD |
| C. necator | VT----- | DSNVVAQYPNLLI | GVVPIVNWLEWIQ | KLWRDKKQ | RGYTEAATD | TLRSMDD |
| R. rubrum | VT----- | DTVVAQHADLKI | GVVPIVNWLEWIQ | LHRRDRAARGY | TEAATD | TLRSMDD |
| R. sphaeroides | VN----- | SEVNIAGLADLKI | GVVPIVNWLEWIQ | KIHRDRAATRGY | TEAATD | TLRSMDD |
| A. cryptum | RH----- | GDIDTGRHADVKI | GVVPIVNWLEWIQ | KIHRDRAARGY | TEAATD | TLRSMDD |
| X. flavus | VT----- | DELNLAQHADLKI | GVVPIVNWLEWIQ | KIHRDKAATRGY | TEAATD | TLRSMDD |
| N. hamburgensis | VT----- | DKVNVAQYADLKI | GVVPIVNWLEWIQ | LHRRDRAARGY | TEAATD | TLRSMDD |
| N. vulgaris | VT----- | DKVNVAQYADLKI | GVVPIVNWLEWIQ | LHRRDRAARGY | TEAATD | TLRSMDD |
| R. palustris DX1 | IT----- | EKVNVAQHADLKI | GVVPIVNWLEWIQ | LHRRDRAARGY | TEAATD | TLRSMDD |
| R. palustris Bis | MT----- | ETVNVAQHADLKI | GVVPIVNWLEWIQ | LHRRDRAARGY | TEAATD | TLRSMDD |
| M. capsulatus | IT----- | DRINVRKYVDLLV | GVVPIVNWLEWIQ | KIHRDRAARGY | TEAATD | TLRSMDD |
| A. ferrooxidans | KT----- | GAVDVTNYVDLLV | GVVPIVNWLEWIQ | KIHRDRAARGY | TEAATD | TLRSMDD |
| C. M. oxyfera | KDISPDQYGGYD | VQYVDLGI | GVVPIVNWLEWIQ | KIHRDRAARGY | TEAATD | TLRSMDD |
| T. denitrificans | KT----- | DTVVAQHVDLGI | GVVPIVNWLEWIQ | KIHRDRAARGY | TEAATD | TLRSMDD |
| A. vinosum | VT----- | EDSNVAQHVDLGI | GVVPIVNWLEWIQ | KIHRDRAARGY | TEAATD | TLRSMDD |
| P. lunula | DE----- | KASAQLDLGI | YDIDVNDVFAW | VQRDVAERGWTE | EQVRAD | DKLRPDD |
| L. polyedrum | DK----- | KASAQLDLGI | YDIDVNDVFAW | VQRDVAERGWTE | EQVRAD | DKLRPDD |
| B. natans | DK----- | DVIESLDFTFY | IDVSDPVKAWK | TEEDMVERG | HKKEDI | IASAESRPDD |
| E. gracilis | DD----- | RVAGLLDFSI | YLDISDRVFAWK | IQDMDAERGH | WALEDI | KKDEKRPDD |
| P. parvum | DE----- | RVNKALDLTVY | LIDITDEVFAW | AQRDIAER | GATMBE | VQKADGKRPDD |
| G. theta | DK----- | RVEELDFSI | YLDITPEVFNW | VQRDHEERGH | LESIKQ | QEAERRPDD |
| T. pseudonana | DE----- | RVELLDLDFSI | YLDISDPVFNW | IQDMDAERGH | LESIMAS | EAERRPDD |
| O. sinensis | DE----- | RVDLLDFSLY | LIDISDEVFNW | IQDMDAERGH | LESILAS | EAERRPDD |
| P. tricorutum | DK----- | RVDLLDFSLY | LIDISDPVFNW | IQDMDAERGH | LESILAS | EAERRPDD |
| D. lutheri | DD----- | KVESMLDLSI | YLDISGVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| O. tauri | DT----- | RVRDMDFDKI | YLDISDDVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| M. commoda | DE----- | RVRDMDFDKI | YLDISDDVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| P. sitchensis | DA----- | RVELLDLDFSI | YLDISNEVFAWK | IQDMDAERGH | LESIKAS | QAERRPDD |
| P. patens | DE----- | RVELLDLDFSI | YLDISDDVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| S. oleracea | DA----- | RVELLDLDFSI | YLDISNEVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| A. thaliana | DE----- | RVDLLDFSLY | LIDISNEVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| S. moellendorf. | DS----- | RVELLDLDFSI | YLDISDAVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| T. aestivum | DE----- | RVELLDLDFSI | YLDISNEVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| P. trichocarpa | DQ----- | RVDLLDFSLY | LIDISNEVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| M. crystallinum | DS----- | RVELLDLDFSI | YLDISNEVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| P. sativum | DS----- | RVELLDLDFSI | YLDISNEVFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| C. variabilis | DE----- | RVNELDFRI | YLDISDEIFAWK | IQDMDAERGH | LESIKAS | EAERRPDD |
| C. reinhardtii | DK----- | RVAELLDLDFSI | YLDISDDIFAWK | IQDMDAERGH | LESIKSS | EAERRPDD |
| V. carteri | DK----- | RVADLLDFSI | YLDISDDIFAWK | IQDMDAERGH | LESIAKSS | EAERRPDD |
| G. sulphuraria | DA----- | RMKQLDFTVY | LIDISDEVFAWK | IQDMDAERGH | KLLENILAS | EAERRPDD |
| G. kilauensis | DK----- | RVDLLDFKVI | YDLSDPITIQWK | IQDMDAERGH | YDDY | IRAESRPDD |
| G. violaceus | DA----- | RVRNLFDFSI | YDLSDPITIQWK | IQDMDAERGH | YEDY | ILKQEAERRPDD |
| T. elongatus | DE----- | RVSLLDLDFSVY | LIDISDDVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| Synechocystis sp | DE----- | RVRELVDLDFSVY | LIDISSEVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| M. aeruginosa | DE----- | RVSLLDLDFSVY | LIDISDEVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| Cyanotheca sp | DE----- | RVSLLDLDFSVY | LIDISDEVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| S. elongatus | DE----- | RVRELVDLDFSVY | LIDISDEVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| A. variabilis | DE----- | RVSLLDLDFSVY | LIDISDEVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| N. spumigena | DE----- | RVSLLDLDFSVY | LIDISDEVFAWK | IQDMDAERGH | YEDY | ILASNAERRPDD |
| A. boonei | DE----- | LRNYLDFKIV | YDPSKDI | WLVKIKRDVEE | RGYKKE | DIKEIRARBPDD |
| F. placidus | DG----- | IBEILDYKIF | VDPARIIRKWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| A. profundus | DG----- | IRDYIDKIF | VDPARIIRLWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| A. veneficus | DG----- | LRNYLDFKIV | YDPSKDI | WLVKIKRDVEE | RGYKKE | DIKEIRARBPDD |
| M. harundinacea | TQ----- | KLREVDLDFKIV | YDPSRAVRRWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. concilii | TE----- | RLASQIDKIF | VDPSSRSVRLWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. thermophila | TE----- | ELRKLSDKIF | VDPSSRSVRRWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. hungatei | TK----- | SLRKYDYTF | VDPERDVYDK | IKRDMKRGYRKE | EELEE | IQRESDD |
| Methanolinea sp | TP----- | GLRDLDFSLY | LIDPASAVREWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. boonei | TP----- | ALRDLDFSVY | LIDPASAVREWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. limicola | TP----- | KLKSLTDFSI | YLDNFTNVAWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. petrolearia | TE----- | KLRELDLDFSI | YLDNFTNVAWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. palustris | TP----- | ALRDLDFSVY | LIDPASAVREWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. marisnigri | TG----- | TLRDRIDKIV | YDPPDPVRAWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| M. liminatans | TP----- | ELRDLDFSLY | LIDPASAVREWK | IKRDMKRGYRKE | EELEE | IQRESDD |
| <i>Gblocks</i> | | | | | | |
| Annotation | | | # | + | # | |

| | 430 | 440 | 450 | 460 | 470 | 480 |
|-----------------------|--------|-------------------------|-----|--------------------------|--------------------|----------------|
| P. profundum | RFRG | -----IEN---VDF | P | ---YLLSMIQGSFMSRHNTLVVPG | GKMS | ---- |
| P. luminescens | RFRD | -----LTQ---IDF | P | ---YLLAMLQGSFVSSINTIVVPG | GKMG | ---- |
| E. amylovora | HFQG | -----LED---IDF | P | ---YLLSMLQGSFISHIKTLVVPG | GKMG | ---- |
| S. flexneri | HFRN | -----LEG---IDF | P | ---WLLAMLQGSFISHINTLVVPG | GKMG | ---- |
| S. medicae | RFAK | -----PQS---IDF | P | ---YLLSMLHNSYMSRAN | IIVVPGDKLD | ---- |
| R. meliloti | RFAK | -----PQS---IDF | P | ---YLLSMLHNSYMSRAN | IIVVPGDKLD | ---- |
| C. necator | RFXN | -----PKG---IDF | P | ---YLLSMIHDSFMSRANTIVVPG | GKME | ---- |
| R. rubrum | RFRD | -----PKG---IDF | P | ---YLLNMLNDSFMSRPNTIVVPG | GKME | ---- |
| R. sphaeroides | RFRN | -----PRG---IDF | P | ---YLTSMIHGSWMSRAN | IIVVPGNKLD | ---- |
| A. cryptum | RFRD | -----PHG---IDF | P | ---YLLAMLHGSFMSRAN | IIVVPGNKFD | ---- |
| X. flavus | RFRD | -----PHG---IDF | P | ---YLLSMIHNSFMSRAN | IIVVPGNKQD | ---- |
| N. hamburgensis | RFXN | -----PRG---IDF | P | ---YLLSMIPNSFMSRAN | IIVVPGSKMD | ---- |
| N. vulgaris | RFXN | -----PRG---IDF | P | ---YLLSMIPSSFMSRAN | IIVVPGSKLD | ---- |
| R. palustris DX1 | RFXN | -----PRG---IDF | P | ---YLLSMIQGSFMSRAN | IIVVPGSKLD | ---- |
| R. palustris Bis | RFXS | -----PRG---IDF | P | ---YLLSMIQGSFMSRAN | IIVVPGSKMD | ---- |
| M. capsulatus | RFXE | -----PGKFV-VDF | P | ---YLLAMLQNSFMSRHNTIVVPG | GKMG | ---- |
| A. ferrooxidans | RFRD | -----PKE---ENF | P | ---TLLQMLPGSFMSRSNTLVVPG | GKMG | ---- |
| C. M. oxyfera | RFXD | -----PKRFV-VDF | P | ---TLLVMINGSFISRNTIVVPG | GKMG | ---- |
| T. denitrificans | RFRN | -----PQG---VDF | P | ---YLLNMLCNSFMSRRNTIVVPG | GKMG | ---- |
| A. vinosum | RFXD | -----PKKLQ-IDF | P | ---YLLSMIHDSFMSRRN | IIVVPGGKMG | ---- |
| P. lunula | SIKK | -----DLSLT-GSK | P | ---GATMKMYDDDF | NPVTVVEMDGEIDMDNMA | ---- |
| L. polyedrum | SIKK | -----DLTLT-GSK | P | ---GATLKMYYDDDF | NAVTVVEMDGEIDMDNME | ---- |
| B. natans | YIME | EGSTVDWDPCAGPGAMA-CPY | P | ---GTRVRYNEMSGEKHAHV | LEVDGVFG | ---ET |
| E. gracilis | YIEK | G-SSVTWKPC--GDNLQ-CEY | P | ---GLQLAYYTEEYM | HPAEVL | EMDGVVH---NL |
| P. parvum | YIMD | EG-ASITWKPN--PNKLT-TSA | P | ---GVLFSYQDEWF | QSVV | LEMDGKID---SL |
| G. theta | YIWD | EG-SDIEWVPP--RNKLA-SSA | P | ---GAGLKIYQKTEKWA | KDAAVIGMDGKYD | ---KI |
| T. pseudonana | YIFD | EG-SEIEWAPS--ADKLS-SPA | P | ---GKLSYKQEYF | ADVAVVEMD | GTFD---NI |
| O. sinensis | YIFD | EG-STIAWTPA--PSKLS-SSG | P | ---GLTMAYGTEDYY | KPAQVEMD | GTFD---NI |
| P. tricorutum | YIFD | EG-SSIEWTPA--PTKLS-SPA | P | ---GKLLAYPEEFF | KDAQV | LEMDGNFD---NI |
| D. lutheri | YIFD | EG-STIEWTPC--GKFLT-CAY | P | ---GKFRYGTEMYM | SEVTV | EMDGRFD---KL |
| O. tauri | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPDTYY | KEVTV | EMDQDF---KL |
| M. commoda | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPDTFF | EEV | VLEMDGQDF---KL |
| P. sitchensis | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPDIYDNEV | VLEMDGQDF | ---RL |
| P. patens | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPDTYY | NEV | VLEMDGQDF---KL |
| S. oleracea | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFSYGPDTFY | NEVTV | VEMDGMFD---RL |
| A. thaliana | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFNYPDSYFDHEV | VLEMDGQDF | ---RL |
| S. moellendorf. | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPDTYYDNEV | VLEMDGQDF | ---KL |
| T. aestivum | YIFD | EG-STINWIPC--GRKLT-CSY | P | ---GKFSYGPDTFY | QEV | VLEMDGQDF---RL |
| P. trichocarpa | YIFD | EG-SSISWIPC--GRKLT-CSY | P | ---GKFSYGPDAYY | HEV | VLEMDGQDF---RL |
| M. crystallinum | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPDTFY | NEVTV | VEMDQDF---RL |
| P. sativum | YIFD | EG-STISWIPC--GRKLT-CSY | P | ---GKFFYGPETYK | NEV | VVEMDQDF---RL |
| C. variabilis | YIFD | QG-STVSWIPC--GRKLT-CSF | P | ---GKMFYGPDTYY | EEV | VLEMDGQDF---KL |
| C. reinhardtii | YIFD | EG-STISWIPC--GRKLT-CSF | P | ---GKMFYGPDTWY | QEV | VLEMDGQDF---KL |
| V. carteri | YIFD | EG-STISWIPC--GRKLT-CSF | P | ---GKMFYGPDTWY | QEV | VLEMDGQDF---KL |
| G. sulphuraria | YIFD | EG-STIDWIPC--GRKLT-CSY | P | ---GKPHYGPDNWNHVDV | LEVDGNFE | ---KL |
| G. kilaeensis | YIFD | RAISDVCW----KPNFP-NSD | P | ---ELTFCYGTDDYF | RPA | YLSIDGEFA---P |
| G. violaceus | YIFD | RAISDVCWK----PNFPN-SDEE | P | ---LTFYCYGTQDYF | RPA | YLSIDGEFA---P |
| T. elongatus | YIFD | EG-STITWIPC--GRKLT-CSY | P | ---GIRLSYGPDEYY | HPV | VLEVDGRFE---KL |
| Synechocystis sp | YIFD | EG-STIDWRPC--GRKLT-CTY | P | ---GKMYGPDNFM | NEV | LEVDGRFE---NL |
| M. aeruginosa | YIFD | EG-STIDWRPC--GRKLT-CTY | P | ---GKLYGPDGFL | NEV | LEVDGQDF---NL |
| Cyanotheca sp | YIFD | EG-STIDWRPC--GRKLT-CAY | P | ---GLKMYGPDNIF | NEV | LEVDGQDF---NL |
| S. elongatus | YIFD | EG-STIQWTPC--GRKLT-CSY | P | ---GIRLAYGPDYY | HEV | VLEVDGQFE---NL |
| A. variabilis | YIFD | EG-STINWTPC--GRKLT-CSY | P | ---GMQLYYGSDVYY | RYV | VLEVDGQDF---NL |
| N. spumigena | YIFD | EG-STINWTPC--GRKLT-CSY | P | ---GMQVYYGSDVYY | RYV | VLEVDGQDF---NL |
| A. boonei | SIINT | -----SQR | P | ---MMISYRDDFYMKRV | RTITDGLIP | ---- |
| F. placidus | TINVDL | -----SRLIK-ASER | P | ---DFAISFFSDYYY | KRA | FIDIDGLFLN---- |
| A. profundus | SINIDI | -----SDLVK-ASER | P | ---DFSIGFFSDYYYAEKA | FIDIDGLFLN | ---- |
| A. veneficus | KIDIDL | -----SAFVR-ASEK | P | ---DFALGFFSDYYYEKPA | FIDIDGLMN | ---- |
| M. harundinacea | DIKIDL | -----SAL---FSL | P | ---DGEFSISFQRDDYY | KRV | VLEVDGELH----R |
| M. concilii | SMNIDL | -----SKILR-RSEH | P | ---EFSIEFQRDDYY | KRVGIMTM | GEIH---- |
| M. thermophila | DITIDL | -----SRIMR-LTER | P | ---EFSIEFQRDDYY | KRVGIMTLD | GEFP---L |
| M. hungatei | ENIDL | -----CDLTK-KSSH | P | ---DFSLSCSISHTPDSRNMRAL | VVDGELM | ---- |
| Methanolinea sp | CGMDL | -----VSLMS-SFDH | P | ---NFMVEYRVVTEA | RRMGR | LAFDGGIP---- |
| M. boonei | SISIDL | -----TSLLA-FHES | P | ---EFSIEFSTQRIECSFLRSL | TFDGE MN | ---- |
| M. limicola | CINFDL | -----FAINS-LADR | P | ---NFRFDFRVTERG | EKIGALS | IDEFQ---Y |
| M. petrolearia | NINFDL | -----FAINS-LAER | P | ---GFSDFSIIEKYEKMGAL | SIDGFR | ---- |
| M. palustris | DISIDL | -----FSLLS-LSDR | P | ---NFLIEFSHEQRNDERTGEL | IILGELS | ---- |
| M. marisnigri | DISIDL | -----FGLLS-LSER | P | ---DFMVEFTIEDVG | EAMGAL | TFDELN---- |
| M. liminatans | GISIDL | -----GEIFRLCER | P | ---FLLEFGLSSLD | REL | ALVLDGELA---- |

Gblocks
Annotation

* *
C243 C249

| | 490 | 500 | 510 | 520 | 530 | 540 |
|-----------------------|--------------------------------------|----------------|------------|----------|---------------|-----------------------------------|
| | =====+=====+=====+=====+=====+=====+ | | | | | |
| P. profundum | ----- | ----- | ----- | FAME | IIVRPLIQQLIDT | GKVG--- |
| P. luminescens | ----- | ----- | ----- | LAME | IIIMPLVQRLL | EGGKIS--- |
| E. amylovora | ----- | ----- | ----- | LAME | IIIMGPLVKRLL | IEGKKIG--- |
| S. flexneri | ----- | ----- | ----- | LAME | IIIMLPLVQRLL | MEGKKIE--- |
| S. medicae | ----- | ----- | ----- | LAMQ | IIIFPLIHLL | LERKHRMS--- |
| R. meliloti | ----- | ----- | ----- | LAMQ | IIIFPLIHLL | LERKHRMS--- |
| C. necator | ----- | ----- | ----- | LAMQ | IIIFPFVLRMM | ERRKRAAQ--- |
| R. rubrum | ----- | ----- | ----- | LSMQ | IIIFPFVLRMM | ERRKRAALGR--- |
| R. sphaeroides | ----- | ----- | ----- | LAMQ | IIILPLIDRVV | VRESKVA--- |
| A. cryptum | ----- | ----- | ----- | LAMQ | IIILPIIMQLM | DRKRRAQ--- |
| X. flavus | ----- | ----- | ----- | LAMQ | IIILPLIMQLM | DRKRRAQ--- |
| N. hamburgensis | ----- | ----- | ----- | LAMQ | IIILPLILQLI | DRKRRA--- |
| N. vulgaris | ----- | ----- | ----- | LAMQ | IIILPLILQLI | ERKKRA--- |
| R. palustris DX1 | ----- | ----- | ----- | LAMQ | IIILPLIMQLI | DRKRSVK--- |
| R. palustris Bis | ----- | ----- | ----- | LAMQ | IIILPLIMQLI | ERKRGA--- |
| M. capsulatus | ----- | ----- | ----- | LAME | IIIFRPILERM | MADREKG--- |
| A. ferrooxidans | ----- | ----- | ----- | YAME | IIILGPRIERM | LDMHLTI--- |
| C. M. oxyfera | ----- | ----- | ----- | LAME | IIILAPIVH | IMIENLKA--- |
| T. denitrificans | ----- | ----- | ----- | FAME | IIILAPLIQDM | IANKKK--- |
| A. vinosum | ----- | ----- | ----- | FAME | IILOPIIERM | MMDARV--- |
| P. lunula | DQLKEIEENLEGLAGS-- | PGELTEAMTKLRSS | PGSON | GTGL | IQVIAM | KVREVEYERLTAKV |
| L. polyedrum | AQLKEIEDNLEGLPSKTP | PGELTEAMVKLKSS | PGSON | GTGL | MIQVIAM | KFREVEYEKLTGGK |
| B. natans | EELFFIEERLSNNTNTKY | FGELTKQMLKNA | APGSD | SL | IQALVAL | KMRESYERFTGRT |
| E. gracilis | KEGLYVEKFLHNTGAKE | FGELTQELLKQNS | PGD | GTGL | FMQLAAL | KIREIYERATGE- |
| P. parvum | EELIYVESQLCNTGT | KYYGELTEQM | VNNKAS | PGSEN | SL | IQICAFKIREAFEAITA-- |
| G. theta | DEMYYVEKQFASTGSK | FVGEITKKMLEY | EGQPGS | ND | GTGL | FLQITALKVREVEYEGIAVKV |
| T. pseudonana | QELVYVESNLGNTNSK | FYGEVTQAMLS | LADSPGS | NN | GTGL | IMQLAFAFAIRELYNKKSAAA |
| O. sinensis | QELVYVESQLSNTSTK | FYGELTQAMLK | LADAPGS | NN | GTGL | IMQLAFAFAIRELYEKKAAAA |
| P. tricornutum | QELVYVESALSNTTK | FYGEMTQAML | LALATAPGS | NN | GTGL | IMQLAFAFAIRDIYEKKTAAA |
| D. lutheri | DELIYVESALNTGAK | FYGEITQILKN | KDAVGS | DN | GTGL | FFQLCSFKIREAYEKATGKT |
| O. tauri | EELIYVESHLNNTSS | KFYGEITQQML | KYQNGP | GSNN | GTGL | FFQIVGLKIREVEYERISKE |
| M. commoda | EELIYVESHLNNTST | KFYGEITQQML | KYQNGP | GSNN | GTGL | FFQIVGLKIREVEYERISEKE |
| P. sitchensis | EELIYVESHLNNTST | KFYGEVTQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRDVYEQILISQT |
| P. patens | DELIYVESHLNNTST | KFYGEITQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRSVYERILANQ |
| S. oleracea | DELIYVESHLNNTST | KFYGEVTQQML | KHQNF | PGSNN | GTGL | FFQIVGLKIRDLEFQIVASR |
| A. thaliana | DELIYVESHLNNTST | KFYGEVTQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRDLYEQLIANK |
| S. moellendorf. | DELIYVESHLNNTST | KFYGEITQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRDVFERITASKQ |
| T. aestivum | DELIYVESHLNNTST | KFYGEVTQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRDLYEQIIAER |
| P. trichocarpa | DELIYVESHLNNTST | KFYGEVTQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRDLEFQIVASR |
| M. crystallinum | DELIYVESHLNNTST | KFYGEVTQQML | KHQDF | PGSNN | GTGL | FFQIVGLKIRDLEFQLIASK |
| P. sativum | DELIYVESHLNNTSS | KFYGEVTQQML | KHADFP | GSNN | GTGL | FFQIVGLKIRDLEFQIVAS- |
| C. variabilis | EELIYVESHLNNTSA | KFYGEITQQML | KNSFP | GSNN | GTGL | FFQIVGLKIREVEYERITQK- |
| C. reinhardtii | EELIYVESHLNNTSA | KFYGEITQQML | KNSGFP | GSNN | GTGL | FFQIVGLKIREVEYERIVKKD |
| V. carteri | EELIYVESHLNNTSA | KFYGEITQQML | KNSGFP | GSNN | GTGL | FFQIVGLKIREVEYERIVKKD |
| G. sulphuraria | EELIYIESHLNNTST | KFYGEITQQL | RNSAPGS | NN | GTGL | FFQIVLTKMRQLYEELTQKS |
| G. kilaeensis | DDSPLEEQKLRNTSE | TRFGEASLIL | KHTDYP | GSRD | GTGL | FFQIVLGLKFRSIEALIAQS |
| G. violaceus | EDSPELEEKLRNTSE | TRFGEASLIL | KHTDYP | GSRD | GTGL | FFQIVLMSLKFRAIYEALTAQL |
| T. elongatus | DELIYIESHLNNTST | KHYGEVTELL | LKHHDYP | GSND | GTGL | FFQIVLTKLMRATYERLTSRD |
| Synechocystis sp | EEMVYVENHLSKTGT | KYYGEMTELL | LKHHDYP | PGTD | GTGL | FFQIVLGLKMRKVYEQLTAAE |
| M. aeruginosa | EEMIIYIESHLSKTG | KYYGEMTELL | LKHHDYP | PGST | GTGL | FFQIVLGLKMRRETYEKLMAAE |
| Cyanothecae sp | EEMIIYIEQHLSTGT | KYYGEMTHLL | QHKDYP | PGSNN | GTGL | FFQIVLGLKMRRETYEILTSV- |
| S. elongatus | EEMIIYVEGHLSKTD | TQYGEITHL | LQHKDYP | PGSNN | GTGL | FFQIVLTKLMRAAYERLTSQ- |
| A. variabilis | EEVIYIETHLSNTST | KYQELTQLL | LQHREYP | GSNN | GTGL | FFQIVLTKLMRAAYERLTAKE |
| N. spumigena | DEVIYVETHLSKTST | KYEGEMTHLL | LQHREYP | GSNN | GTGL | FFQIVLTKLMRAAYERLTTK- |
| A. boonei | ----- | HEAIDSLERRIME | YTGFFENY | ILERSKY | VNG | QIALLVAVYFVEMMNIFREI |
| F. placidus | ---VEIFESLLDALR | DETYKVRW---- | EAREYVNS | IEVAKLLC | WNLV | EIIKMKG--- |
| A. profundus | ---VDIFKSLFD | SLRKEIGD---- | EIKVESEY | VNAIEF | SKLL | VWCMLVEVLKHSR-- |
| A. veneficus | ---VELFSTLLD | LKKEVGAD | VEWKMT---- | YVNAIE | VAKLL | VWCWFLELIKFRLEAE |
| M. harundinacea | SMVEGVEAKL | KSLAG----- | STAPIC | DRAGD | RVT | SGMAQLLSWRLLKQLHLDKA |
| M. concilii | ----- | QSMISDLEK | KLGS | ELGTD | GMSDR | REYVNAIEALILTWNCVEKLDYLLQEE |
| M. thermophila | TMIKDLERKLCDF | LGRDVP | SVTEG-- | CQED | HGSY | VNAIEMTQLILIRWFLEKIANLAW-- |
| M. hungatei | ----- | PDTIHKIERQIE | FQTGIS | PINIFR | QEHIT | STDVRLILSWQIINGRIALS NHL |
| Methanolinea sp | ----- | REMAGDLRR | KIEEQ | TGTQ | PADM | FRGNAMVTAIDMVRLIVSWRIINHLANLA--- |
| M. boonei | ----- | YDTRVNR | LELSIE | HQTGI | HPIEM | FAGRKVVTPINIVQLLSWRIINRRIFLQDHR |
| M. limicola | ----- | HEVVSLEK | GIEM | QTGIG | PVSV | SDRSYVTAEMIVQLLSWRIINRRIMQES-- |
| M. petrolearia | ----- | HEVVSLLEK | GIEM | QTGIG | PVSV | SDRSYVTAEMIVQLLSWRIINRRIMQES-- |
| M. palustris | ----- | TRMVKRL | ETSIEE | QTQ | VRPIS | DFHDHYMTAEVVLILAWRIHQRVFLERCL |
| M. marisnigri | ----- | DAVARKL | ERNIE | IQTQ | VEPID | LSQSDSYLTAGDMAQLILAWRIINRRIFIESAP |
| M. liminatans | ----- | ASAIRRL | ARTIG | RETR | SEAVN | LADRQYVTAGEIAELILAWRIINRWHVLEAAR |
| Gblocks | | | | | | |
| Annotation | | | | | | |

| | 550 | 560 |
|-----------------------|---------------------|-------|
| | =====+=====+ | |
| P. profundum | ----- | ----- |
| P. luminescens | ----- | ----- |
| E. amylovora | ----- | ----- |
| S. flexneri | ----- | ----- |
| S. medicae | ----- | ----- |
| R. meliloti | ----- | ----- |
| C. necator | ----- | ----- |
| R. rubrum | ----- | ----- |
| R. sphaeroides | ----- | ----- |
| A. cryptum | ----- | ----- |
| X. flavus | ----- | ----- |
| N. hamburgensis | ----- | ----- |
| N. vulgaris | ----- | ----- |
| R. palustris DX1 | ----- | ----- |
| R. palustris Bis | ----- | ----- |
| M. capsulatus | ----- | ----- |
| A. ferrooxidans | ----- | ----- |
| C. M. oxyfera | ----- | ----- |
| T. denitrificans | ----- | ----- |
| A. vinosum | ----- | ----- |
| P. lunula | GA----- | ----- |
| L. polyedrum | ----- | ----- |
| B. natans | IQIPKWGLFLDDY----- | ----- |
| E. gracilis | KA----- | ----- |
| P. parvum | ----- | ----- |
| G. theta | VPAQAN----- | ----- |
| T. pseudonana | KLAATKETAASA----- | ----- |
| O. sinensis | KLAVIEA----- | ----- |
| P. tricornutum | KAKAGVSAAAA----- | ----- |
| D. lutheri | VDASAAA----- | ----- |
| O. tauri | VVAKA----- | ----- |
| M. commoda | VVTAA----- | ----- |
| P. sitchensis | TGASLEAAKA----- | ----- |
| P. patens | KNAATLQSAKA----- | ----- |
| S. oleracea | STATATAAKA----- | ----- |
| A. thaliana | ATARAEEKA----- | ----- |
| S. moellendorf. | GSPVGAAAATSKV----- | ----- |
| T. aestivum | AGVPAEAAKV----- | ----- |
| P. trichocarpa | AKTPVEATKA----- | ----- |
| M. crystallinum | TAAPAAATKA----- | ----- |
| P. sativum | RAETPVGAACA----- | ----- |
| C. variabilis | ----- | ----- |
| C. reinhardtii | VVPV----- | ----- |
| V. carteri | VVPA----- | ----- |
| G. sulphuraria | IAPVLV----- | ----- |
| G. kilaeensis | LAAAKV----- | ----- |
| G. violaceus | LTASKAK----- | ----- |
| T. elongatus | AATVTNR----- | ----- |
| Synechocystis sp | KVPASV----- | ----- |
| M. aeruginosa | AKVAASV----- | ----- |
| Cyanothece sp | ESKVATQV----- | ----- |
| S. elongatus | AAPVAASV----- | ----- |
| A. variabilis | AKLAVQV----- | ----- |
| N. spumigena | EAKLAVQV----- | ----- |
| A. boonei | ENAREKNIY----- | ----- |
| F. placidus | ----- | ----- |
| A. profundus | ----- | ----- |
| A. veneficus | L----- | ----- |
| M. harundinacea | ----- | ----- |
| M. concilii | GY----- | ----- |
| M. thermophila | ----- | ----- |
| M. hungatei | DQ----- | ----- |
| Methanolinea sp | ----- | ----- |
| M. boonei | ----- | ----- |
| M. limicola | ----- | ----- |
| M. petrolearia | GVSS----- | ----- |
| M. palustris | HQDHNK----- | ----- |
| M. marisnigri | GAGGTGRTVTGNNHGCGRR | ----- |
| M. liminatans | SAHARG----- | ----- |
| Gblocks | | |
| Annotation | | |

B

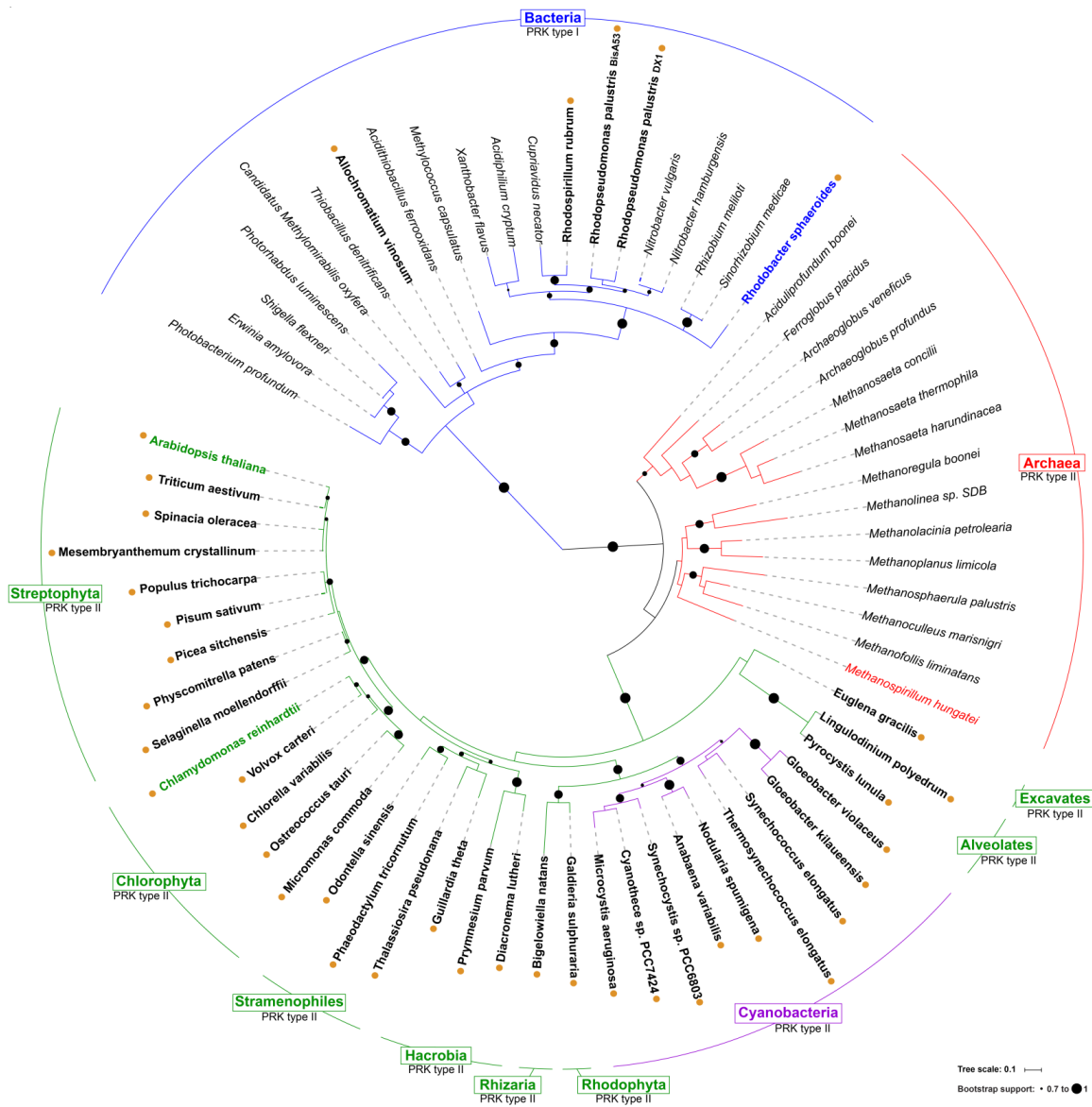


Fig. S6. Sequence alignment and phylogenetic analysis of 69 PRKs. (A) The sequences annotated as PRKs were retrieved from Uniprot and aligned using the phylogeny webserver suite (www.phylogeny.fr) (27). Blue areas in the “Gblocks” line define the conserved areas, later used by the software to determine the phylogeny. The alignment was performed by MUSCLE and curation by Gblocks. Annotations used are as follow: black boxes within the sequences indicate conservation of the residue in more than 70% of the sequences. Walker A (P-loop) and Walker B motives are represented by a red or green area, respectively, and the clamp loop is highlighted by a yellow area. Bars (|) or Hashtags (#) denote residues implicated in Ru5P or ATP binding, respectively. Plus sign (+) indicates two Aspartate residues shown to be crucial for catalysis by mutagenesis in *R. sphaeroides* (28). Star signs (*) indicate Cys residues implicated in disulfide bridges (Cys16 with Cys55; Cys243 with Cys249) (29), numbers below are for *C. reinhardtii* PRK. Blue and red bars on the right side indicate clusters of bacterial and archeal PRK, respectively, while the green one indicates the cluster of eukaryotic and the purple one is for the cyanobacterial PRKs. Species names in bold are indicating the 4 species with known structure. Uniprot accessions numbers and other details, for all protein sequences are reported in Table S6. (B) The phylogeny was built with PhyML and the tree with TreeDyn. The visual was obtained with iTOL (<http://itol.embl.de/>) (30). Bootstrap values superior to 0.7 are represented by black circles with a radius proportional to the value. Branch lengths are represented by straight lines at indicated scale, while dashes are presented for the sake of clarity. The clades are colored in function of their kingdom (Bacteria in blue, Archaea in red and Eukaryotes in green) except Cyanobacteria clade which is in purple. Photosynthetic species are in bold indicated by a yellow circle while the others are italicized. Species for which the PRK has a known structure, *i.e.* *R. sphaeroides* (31), *M. hungatei* (5), *C. reinhardtii* and *A. thaliana*, are represented in the color of their kingdom.

Fig. S7. Sequence and structural alignment of the four structurally known PRKs. The alignment was performed with Esprict (<http://esprict.ibcp.fr>) (32) using the sequence and the structure of *At*PRK (Uniprot accession code P25697 and PDB ID code 6H7H); *Cr*PRK (Uniprot accession code P19824 and PDB ID code 6H7G); *Rs*PRK (Uniprot accession code P12033 and PDB ID code 1A7J) (31), *Mh*PRK (Uniprot accession code Q2FUB5 and PDB ID code 5B3F) (5). The sequence of both photosynthetic PRKs is much longer (349 and 344 residues for *At*PRK and *Cr*PRK, respectively) than bacterial (290 residues) and Archae (323 residues) PRKs. Sequence identities among considered PRKs are: 75% for *At*PRK vs *Cr*PRK; 35% for *At*PRK vs *Mh*PRK; 32% for *Cr*PRK vs *Mh*PRK; 22% for *At*PRK vs *Rs*PRK; 24% for *Cr*PRK vs *Rs*PRK. The sequence identities were calculated by Clustal Omega (33).

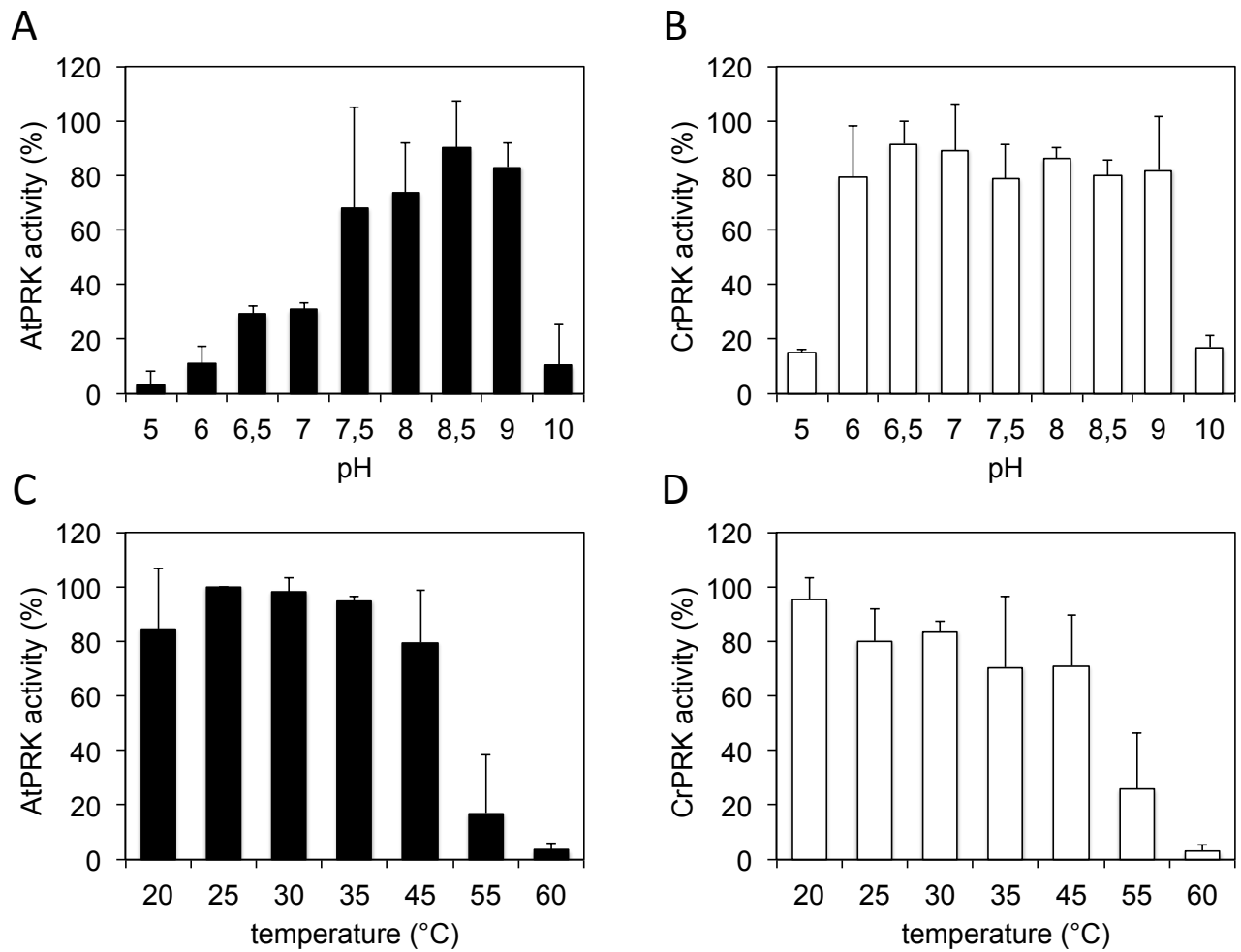


Fig. S8. pH and temperature dependence of photosynthetic PRKs. The enzyme activity of *At*PRK (A and C) and of *Cr*PRK (B and D) was evaluated at different pHs (upper panels) and temperatures (lower panels). Values are reported as means \pm SD (n=3).

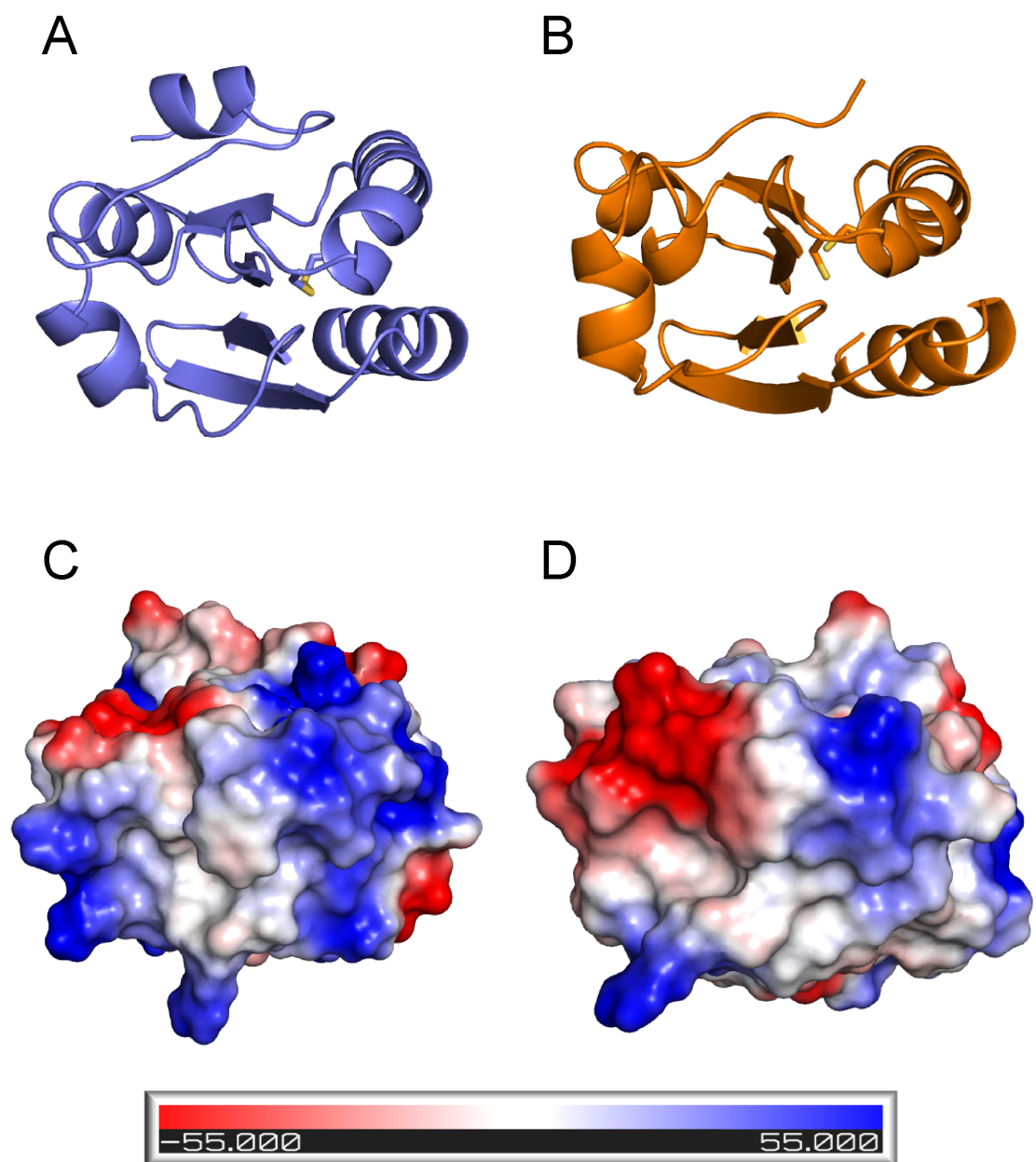


Fig. S9. Electrostatic surface potential of *A. thaliana* TRX-f1 and TRX-m2. Structure represented as ribbon (A, B) and electrostatic surface potential (C, D) of the homology model of *Arabidopsis thaliana* TRX-f1 (A, C) and TRX-m2 (B, D). The crystal structures of *Spinacia oleracea* TRX-f and TRX-m (PDB ID codes 1FAA and 1FB6) (34) were used as template to model *Arabidopsis thaliana* TRX-f1 and TRX-m2, respectively. The sequence identity of *Arabidopsis thaliana* TRXs with spinach enzymes is 59% and 75% for TRX-f1 and TRX-m2, respectively. The homology modelling was performed with Swiss-Model (35).

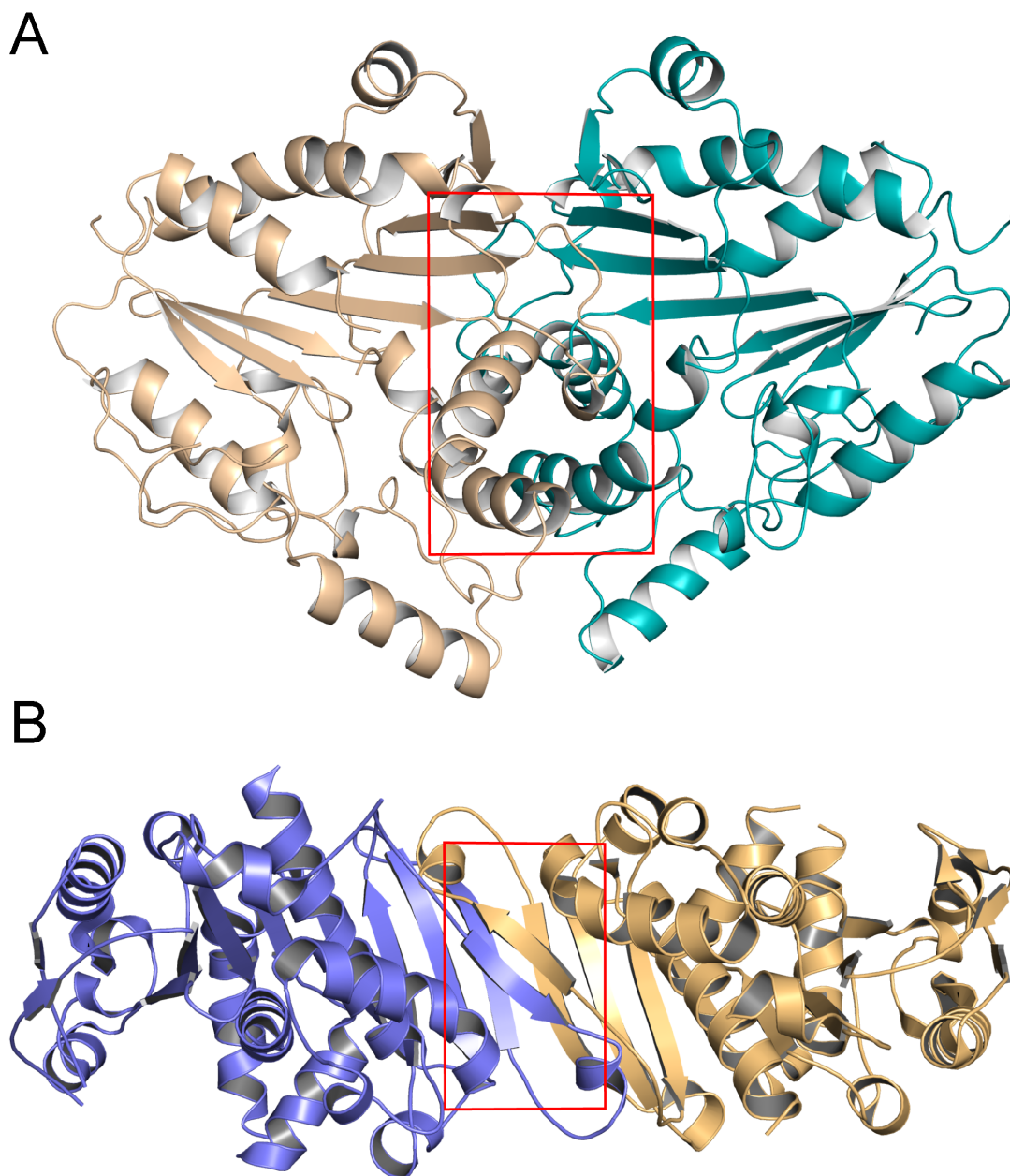


Fig. S10. Dimer interface of bacterial and Archaea PRKs. Dimer interface of (A), octameric *Rhodobacter sphaeroides* PRK (PDB ID code 1A7J) (31) and (B), dimeric *Methanospirillum hungatei* PRK (PDB ID code 5B3F) (5) is highlighted by a red box. The dimer interface of bacterial PRK is formed by three β -strands and one α -helix while in Archaea enzyme by two β -strands. The calculated dimer interface areas are 1667 and 1695 \AA^2 , respectively.

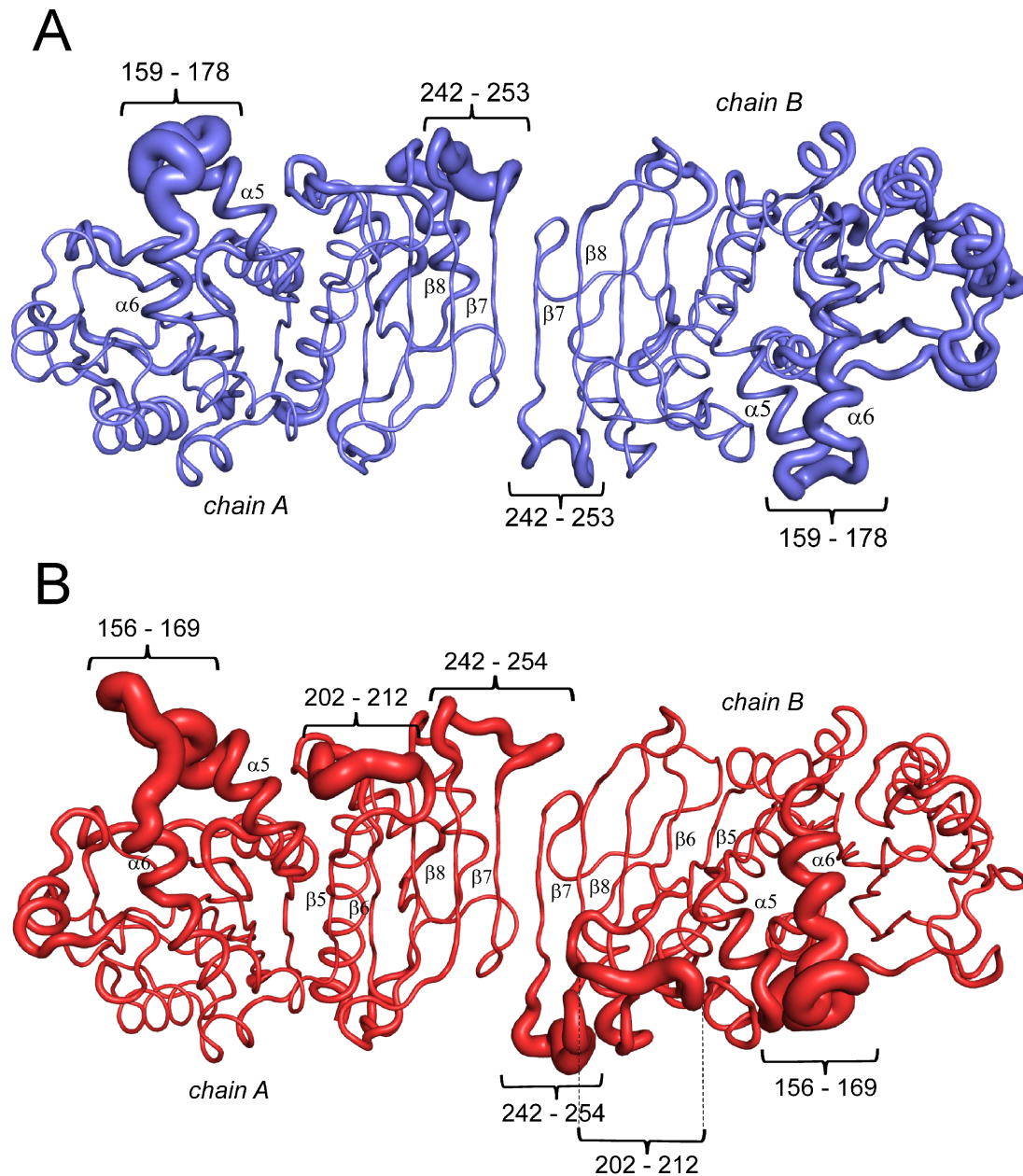


Fig. S11. Flexible and disordered regions in redox-sensitive PRKs. *C.* trace of (A), *CrPRK* and (B), *AtPRK*. The trace thickness is proportional to the atomic B factor. *AtPRK* shows a higher number of flexible regions compared to *CrPRK*. The residues belonging to the flexible regions are reported.

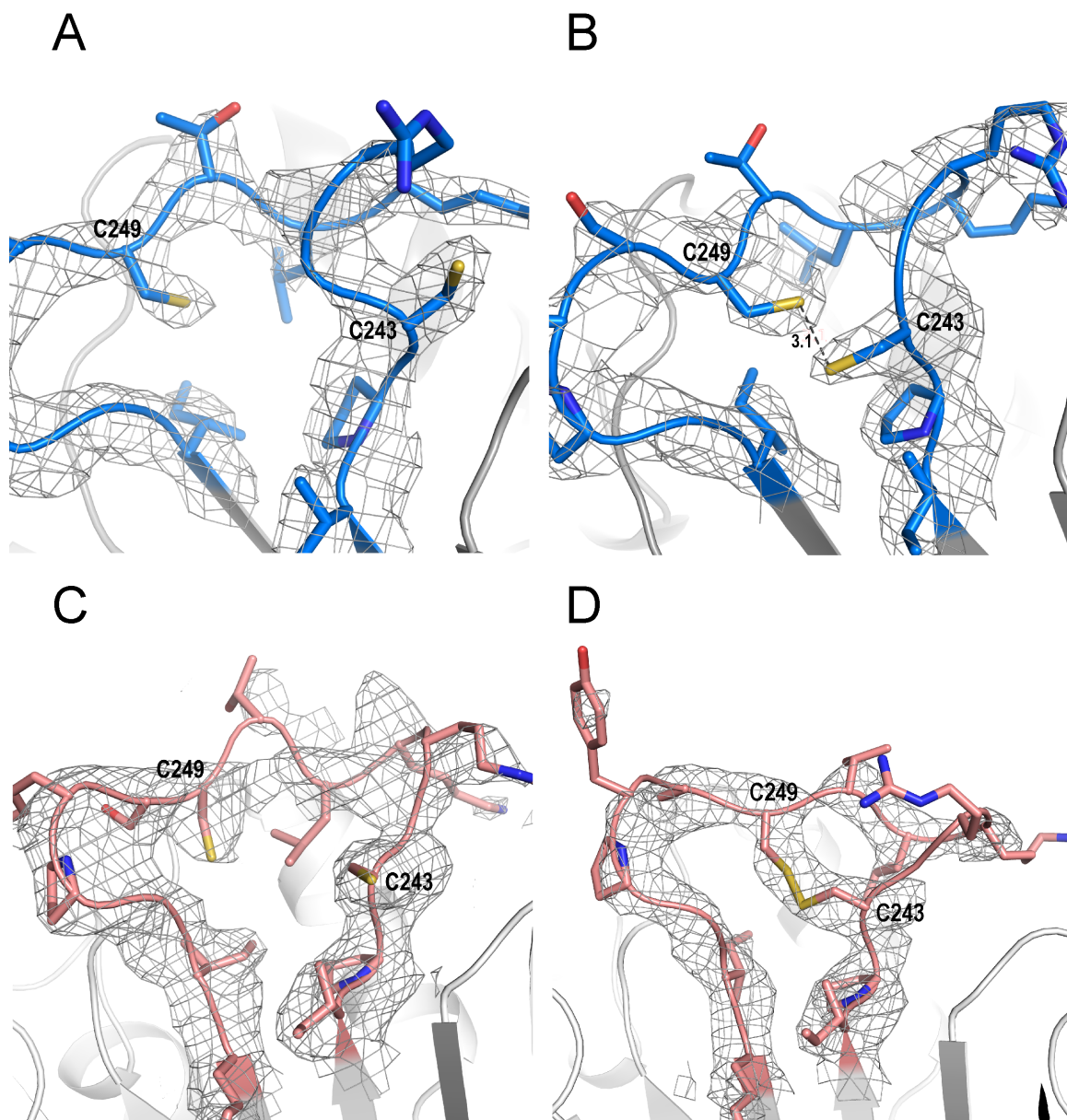


Fig. S12. Electron density of the C-terminal cysteine pair. $2F_o - F_c$ electron density map contoured at 1.2σ and associated with Cys243 and Cys249 in (A), *CrPRK* subunit A (B), *CrPRK* subunit B (C), *AtPRK* subunit A and (D), *AtPRK* subunit B. In *CrPRK*, both C-terminal cysteines are reduced even if in subunit B the thiol groups are only 3 Å apart. A disulfide bond is instead observed in subunits B of *AtPRK*, while the thiol groups of the other cysteine pair (subunit A) lie very distantly.

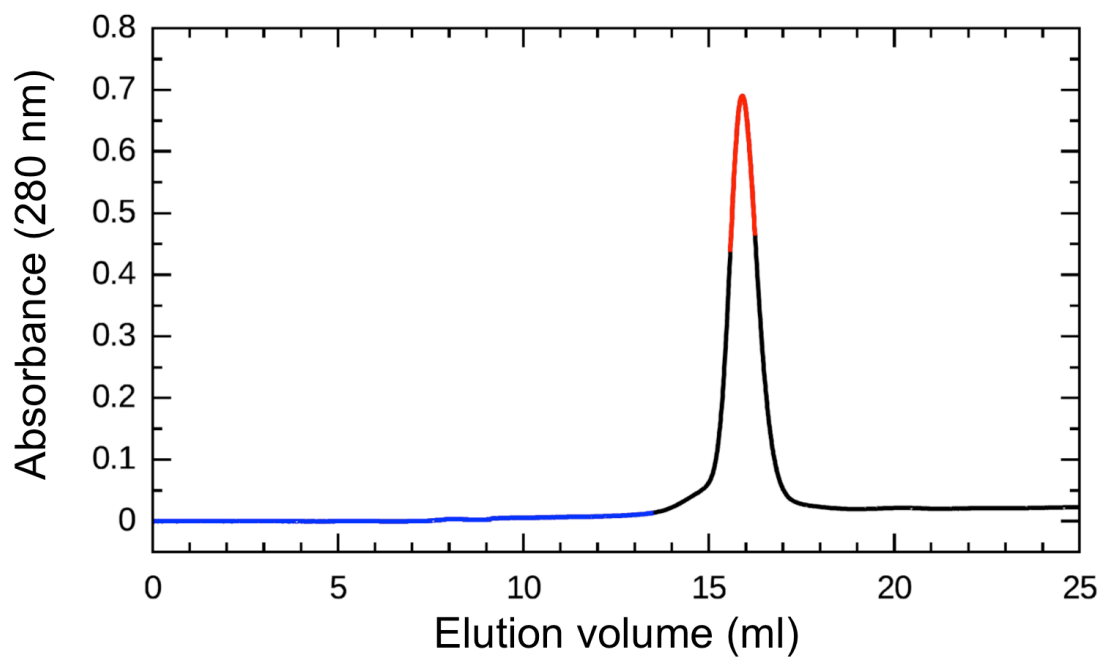


Fig. S13. UV trace of the chromatogram profile of *C7PRK* analyzed in SEC-SAXS mode.

Frames collected as buffer (from 0 to 14 ml) are highlighted in blue; frames collected as protein (from 14 to 17.5 ml) are highlighted in black; frames highlighted in red have been used for SAXS analysis.

Table S1. SEC-SAXS data analysis of reduced CrPRK.

| | |
|---------------------------------------------------------------------------------------------------|----------------|
| Concentration (mg ml ⁻¹) | 6.1 (injected) |
| Structural parameters | |
| q interval for Guinier linear fit (nm ⁻¹) | 0.12-0.38 |
| I(0) [from Guinier approximation] | 42.0 ± 0.1 |
| R _g (nm) [from Guinier approximation] | 3.43 ± 0.01 |
| q interval for Fourier inversion (nm ⁻¹) | 0.12-3.5 |
| I(0) [from P(R)] | 42.4 ± 0.06 |
| R _g (nm) [from P(R)] | 3.55 ± 0.01 |
| D _{max} (nm) | 11.2±0.5 |
| Porod volume estimate (nm ³) | 115 ± 10 |
| DAMMIN excluded volume (nm ³) | 138 ± 1 |
| Dry volume calculated from sequence (nm ³) (v=0.735 cm ³ g ⁻¹) | 95 |
| Molecular mass (kDa) | |
| From I(0) | 70 |
| From Vc | 70 |
| From Porod invariant | 85 |
| From Porod volume (x0.625) | 72 |
| From excluded volume (x0.5) | 69 |
| From sequence | 77.8 |

Table S2. Accessibility values (ASA) for cysteine residues and a strictly conserved arginine.

| Residue (Cr/At) | ASA (\AA^2)* | | | |
|--------------------|-------------------------|------|-------|------|
| | CrPRK | | AtPRK | |
| | A | B | A | B |
| Cys16/15 | 14.5 | 8.5 | 7.6 | 10.6 |
| Cys55/54 | 10.6 | 10.6 | 6.3 | 9.5 |
| Cys61 | 55.9 | 58.7 | / | / |
| Cys243 | 26.6 | 37.8 | 53.4 | 14.8 |
| Cys249 | 42.4 | 31.1 | 14.8 | 50.6 |
| Arg64/63 | 51.4 | 88.7 | 82.6 | 88.2 |

*Radius of the probe solvent molecule 1.4 \AA

Table S3. Secondary structure element content in the structurally known PRKs.

| PRK | Quaternary structure | Helix content (%) | Sheet content (%) | Other (%) |
|-----------------------|----------------------|-------------------|-------------------|-----------|
| <i>C. reinhardtii</i> | Dimer | 31.2 | 21.9 | 46.9 |
| <i>A. thaliana</i> | Dimer | 32.3 | 21.9 | 45.8 |
| <i>M. hungatei</i> | Dimer | 50.0 | 22.2 | 27.8 |
| <i>R. sphaeroides</i> | Octamer | 41.2 | 16.9 | 41.9 |

Table S4. X-ray (CrPRK and AtPRK) and SEC-SAXS (CrPRK) data collection parameters.

| | CrPRK | AtPRK | CrPRK SEC-mode |
|-------------------------------------|------------|--------------------|-------------------|
| Detector | Pilatus 2M | ADSC Quantum Q315r | Pilatus 1M |
| Beam geometry (mm ²) | 0.1 × 0.1 | 0.1 × 0.1 | 0.7 × 0.7 |
| Wavelength (Å) | 1.240 | 0.940 | 0.990 |
| Capillary diameter (mm) | / | / | 1.8 |
| Sample-to-detector Distance (mm) | 239.85 | 393.43 | 2872 |
| $\Delta\phi$ (°) | 0.5 | 0.7 | / |
| q* range (nm ⁻¹) | / | / | 0.033-4.9 |
| Exposure time (s) | 5 | 5 | 1 |
| Flow (ml/min) | / | / | 0.5 |
| Temperature (K) | 100.0 | 100.0 | 277.15 |

*q = $4\pi \sin(\theta)/\lambda$, where 2θ is the scattering angle and λ is the X-ray wavelength.

Table S5. X-ray data collection and refinement statistics.

| | CrPRK | AtPRK |
|-------------------------------------------|-----------------------------------------------|------------------------------------------------|
| <i>Data collection</i> | | |
| Unit cell (Å) | 77.68, 83.55, 133.15, 90.00, 90.00, 90.00 | 116.30, 116.30, 106.81, 90.00, 90.00, 90.00 |
| Space group | P2 ₁ 2 ₁ 2 ₁ | I4 ₁ |
| Resolution range* (Å) | 44.38 – 2.60 (2.72 – 2.60) | 82.23 – 2.47 (2.58 – 2.47) |
| Unique reflections | 27230 (3280) | 24824 (3107) |
| Completeness* (%) | 99.6 (100.0) | 97.6 (99.6) |
| R _{merge} * | 0.076 (0.770) | 0.091 (0.435) |
| CC _{1/2} | 0.997 (0.743) | 0.997 (0.947) |
| I/σ(I) * | 13.6 (1.7) | 11.2 (2.3) |
| Multiplicity* | 5.1 (5.3) | 6.9 (6.5) |
| <i>Refinement</i> | | |
| Resolution range* (Å) | 39.86 – 2.60 (2.69 - 2.60) | 46.77 – 2.47 (2.57 – 2.47) |
| Reflection used | 27162 (2666) | 24672 (2786) |
| R/R _{free} * | 0.227/0.262 | 0.226/0.281 |
| rmsd from ideality (Å, °) | 0.004, 0.915 | 0.011, 1.128 |
| <i>N° atoms</i> | | |
| Non-hydrogen atoms | 5360 | 5385 |
| Protein atoms | 5319 | 5355 |
| Solvent molecules | 31 | 30 |
| Hetero atoms | 10 | / |
| <i>B value (Å²)</i> | | |
| Mean | 62.4 | 78.4 |
| Wilson | 59.0 | 52.5 |
| Protein atoms | 62.3 | 78.4 |
| Solvent molecules | 56.2 | 70.3 |
| Hetero atoms | 85.5 | / |
| <i>Ramachandran plot (%)</i> [§] | | |
| Most favoured | 91.1 | 91.4 |
| Allowed | 7.4 | 6.0 |
| Disallowed | 1.5 | 2.7 |

*Values in parentheses refer to the last resolution shell

[§]As defined by MolProbity (35)

Table S6. Proteins used for phylogeny.

| Organism | Organism - full description | Uniprot used | other Ref. | other Ref 2 | Kingdom | Phylum | Photosynthetic | Reference and/or status of the uniprot entry |
|-----------------------------|-----------------------------------------------------------------------------------------------------------|--------------|------------|-------------|-----------|-------------------|-------------------------|--------------------------------------------------------|
| <i>A. profundus</i> | Archaeoglobus profundus | D2RE99 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) |
| <i>M. concilli</i> | Methanoseta concilli | F4BW53 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) |
| <i>M. hungatei</i> | Methanospirillum hungatei | Q2FUB5 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) |
| <i>M. marisnigri</i> | Methanococcus marisnigri | A3CW00 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) |
| <i>M. thermophila</i> | Methanoseta thermophila | A08947 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) |
| <i>A. boonei</i> | Aciduliprofundum boonei (strain DSM 19572 / 7469) | B5IC08 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>F. placidus</i> | Ferroglobus placidus (strain DSM 11195 / SN96) | FZKMR8 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. boonei</i> | Methanogula boonei (strain DSM 21154 / JCM 14090 / 648) | D3R252 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. horandinaeae</i> | Methanoseta horandinaeae (strain 64c) | A7H16 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. limicola</i> | Methanoplanus limicola DSM 2279 | G7WU21 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. liminatans</i> | Methanofollis liminatans DSM 4140 | JOAT18 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. parvioris</i> | Methanospirillum parvioris (strain ATCC 6AA-1556 / DSM 19558 / E1-9c) | B6G652 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. petrolearius</i> | Methanodanthonia petrolearia (strain DSM 11571 / DSM 11571 / DSM 11571 / DSM 11571 / DSM 11571) | ELN35 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>Methanodanthonia sp.</i> | Methanodanthonia sp. 30b | AF4903115 | | | Archae | Euryarchaeota | no | Kono et al., 2017 (5) / protein predicted |
| <i>A. ferrooxidans</i> | Acidithiobacillus ferrooxidans (strain ATCC 33270 / DSM 14882 / CIP 104768 / NCIMB 8455) | B75555 | | | Bacteria | Acidithiobacillia | no | Kono et al., 2017 (5) / protein inferred from homology |
| <i>R. sphaeroides</i> | Rhodospirillum rubrum | P12033 | | | Bacteria | Acidithiobacillia | no | Hallenbeck and Kaplan, 1987 (36) |
| <i>N. vulgaris</i> | Nitrosobacter vulgaris | AAA226113 | | PDB-1A71 | Bacteria | no | yes (purple non-sulfur) | inferred by homology |
| <i>R. malii</i> | Rhizobium malii (strain 1021) (Einfelder meliloti) (Sinorhizobium meliloti) | P37100 | | | Bacteria | no | no | inferred by homology |
| <i>S. medicae</i> | Sinorhizobium medicae (strain WSM419) (Einfelder medicae) | P58347 | | | Bacteria | no | no | inferred by homology |
| <i>R. pallidus DX</i> | Rhodospirillum rubrum | AUI46366 | | EF6VW3 | Bacteria | no | yes (purple non-sulfur) | inferred by homology |
| <i>R. rubrum</i> | Rhodospirillum rubrum | ABC3204 | | Q2RRP1 | Bacteria | no | yes (purple non-sulfur) | Kono et al., 2017 (5) |
| <i>A. cryptum</i> | Acidiphilium cryptum (strain IF-5) | A5FW09 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein predicted |
| <i>M. hamburgensis</i> | Nitrobacter hamburgensis (strain DSM 10229 / NCIMB 13809 / X14) | Q07N64 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein inferred from homology |
| <i>R. pallidus B1</i> | Rhodospirillum rubrum | P37101 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein inferred from homology |
| <i>X. flavus</i> | Xanthobacter flavus | P37101 | | | Bacteria | no | no | Meijer et al., 1991 (37) |
| <i>Synchocystis sp.</i> | Synchocystis sp. (strain PCC 6803 / Kazusa) | P37101 | | | Bacteria | no | no | Experimental evidence at transcript level |
| <i>S. elongatus</i> | Synchococcus elongatus PCC 7942 | G31P12 | | | Bacteria | Cyanobacteria | yes | Kobayashi et al., 2003 (38) |
| <i>A. variabilis</i> | Anabaena variabilis (strain ATCC 29413 / PCC 7937) | Q3MF31 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein inferred from homology |
| <i>G. veloxus</i> | Gloeobacter veloxus (strain PCC 7421) | Q7N87 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein inferred from homology |
| <i>T. elongatus</i> | Thermosynechococcus elongatus (strain BP-1) | Q8DH2 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein inferred from homology |
| <i>Cyanotheca sp.</i> | Cyanotheca sp. (strain PCC 7424) (Synchococcus sp. (strain ATCC 29155)) | B7K162 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein predicted |
| <i>G. klauaensis</i> | Gloeobacter klauaensis JS1 | U5QBW9 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein predicted |
| <i>M. aeruginosa</i> | Microcystis aeruginosa PCC 9701 | I4IN9 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein predicted |
| <i>N. spumigena</i> | Nodularia spumigena CCY9414 | AG2E1 | | | Bacteria | Cyanobacteria | yes | Kono et al., 2017 (5) / protein predicted |
| <i>E. amylovora</i> | Erwinia amylovora | ESBA03 | | | Bacteria | no | no | inferred by homology |
| <i>S. flexneri</i> | Shigella flexneri | POAEV6 | | | Bacteria | no | no | inferred by homology |
| <i>M. capsulatus</i> | Methylococcus capsulatus (strain ATCC 33009 / NCIMB 11132 / Bath) | Q6Q212 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein predicted |
| <i>A. vinosum</i> | Allochromatium vinosum (strain ATCC 17899 / DSM 180 / NBRC 103801 / NCIMB 10441 / D) (Chromatium vinosum) | D3R02 | | | Bacteria | no | yes (purple sulfur) | Kono et al., 2017 (5) / protein inferred from homology |
| <i>P. luminescens</i> | Photobacterium luminescens (Xenorhabdus luminescens) | AOA188YK5 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein inferred from homology |
| <i>P. profundum</i> | Photobacterium profundum (strain S59) | GLV61 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein inferred from homology |
| <i>C. M. oxyfera</i> | Candidatus Methylohalobium oxyfera | D5MH04 | | | Bacteria | Unclassified | no | Kono et al., 2017 (5) / protein inferred from homology |
| <i>T. denitrificans</i> | Thiobacillus denitrificans (strain ATCC 25259) | C3S556 | | | Bacteria | no | no | Kono et al., 2017 (5) / protein predicted |
| <i>C. neoator</i> | Cupriavidus necator | P19923 | | | Bacteria | no | no | Kossmann et al., 1989 (39) |
| <i>E. gracilis</i> | Euella gracilis | Q24L70 | | | Excavates | Euglenozoa | yes | Experimental evidence at transcript level |
| <i>G. theta</i> | Galliardia theta | Q24L74 | | | Excavates | Cryptophyta | yes | Experimental evidence at transcript level |
| <i>D. lutheri</i> | Dicranema lutheri (Monochrysis lutheri) Pavlova lutheri | Q24L75 | | | Excavates | Haptophyceae | yes | Experimental evidence at transcript level |
| <i>C. variabilis</i> | Chlorella variabilis | Q24L58 | | | Excavates | Haptophyceae | yes | Experimental evidence at transcript level |
| <i>M. commoda</i> | Micromonas commoda | C1FW3 | | | Excavates | Chlorophyte | yes | Predicted protein |
| <i>O. tauri</i> | Ostreococcus tauri | ELF27 | | | Excavates | Chlorophyte | yes | inferred by homology |
| <i>V. carteri</i> | Volvox carteri f. nagariensis | C1FW3 | | | Excavates | Chlorophyte | yes | inferred by homology |
| <i>G. sulphuraria</i> | Gliamydomonas reinhardtii | AA090N369 | | | Excavates | Chlorophyte | yes | inferred by homology |
| <i>P. sativum</i> | Pisum sativum | D8TR87 | | | Excavates | Chlorophyte | yes | Predicted protein |
| <i>P. sativum</i> | Pisum sativum | P19924 | AS1V4 | AAF3642 | Excavates | Rhodophyta | yes | Roester and Ogren, 1990 (40) |
| <i>P. sativum</i> | Pisum sativum | P93661 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>P. sativum</i> | Pisum sativum | CP9566 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>P. sativum</i> | Pisum sativum | AS1V4 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>P. sativum</i> | Pisum sativum | B9Z275 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>P. sativum</i> | Pisum sativum | DSU948 | | | Excavates | Rhodophyta | yes | inferred by homology |
| <i>S. moellendorffii</i> | Sesuvium portulacastrum | P27274 | | | Excavates | Rhodophyta | yes | inferred by homology |
| <i>M. crystallinum</i> | Mesembryanthemum crystallinum | P27274 | | | Excavates | Rhodophyta | yes | inferred by homology |
| <i>Z. mays</i> | Zinnia mays | P27274 | | | Excavates | Rhodophyta | yes | inferred by homology |
| <i>A. asperum</i> | Arabidopsis thaliana | P35559 | | CAA30499 | Excavates | Rhodophyta | yes | Roester and Ogren, 1990 (40) |
| <i>A. thaliana</i> | Arabidopsis thaliana | P35559 | | | Excavates | Rhodophyta | yes | Roester and Ogren, 1990 (40) |
| <i>L. perfoliatus</i> | Lepidodermis perfoliatus | P35559 | | | Excavates | Rhodophyta | yes | Roester and Ogren, 1990 (40) |
| <i>P. lundii</i> | Populus lundii | Q24L73 | | | Excavates | Rhodophyta | yes | This Study and Ogren, 1990 (40) |
| <i>B. nana</i> | Begonia nana | Q24L73 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>B. nana</i> | Begonia nana | Q24L73 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>O. sinensis</i> | Oenothera sinensis | Q24L73 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| <i>P. tricornutum</i> | Phaeodactylum tricornutum | OS033 | | | Excavates | Rhodophyta | yes | Predicted protein |
| <i>T. pseudonana</i> | Thalassiosira pseudonana | B5V5F0 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |
| | | B8B240 | | | Excavates | Rhodophyta | yes | Experimental evidence at transcript level |

**Member of the SAR group (42). #Member of the Hacrobia kingdom (43)

References

1. Kabsch W (2010) XDS. *Acta Crystallogr D Biol Crystallogr* 66(Pt 2):125–132.
2. Evans P (2006) Scaling and assessment of data quality. *Acta Crystallogr D Biol Crystallogr* 62(Pt 1):72–82.
3. McCoy AJ, et al. (2007) Phaser crystallographic software. *J Appl Crystallogr* 40(Pt 4):658–674.
4. Adams PD, et al. (2010) PHENIX: a comprehensive Python-based system for macromolecular structure solution. *Acta Crystallogr D Biol Crystallogr* 66(Pt 2):213–221.
5. Kono T, et al. (2017) A RuBisCO-mediated carbon metabolic pathway in methanogenic archaea. *Nat Commun* 8:14007.
6. Cowtan K (2006) The Buccaneer software for automated model building. 1. Tracing protein chains. *Acta Crystallogr D Biol Crystallogr* 62(Pt 9):1002–1011.
7. Murshudov GN, Vagin AA, Dodson EJ (1997) Refinement of macromolecular structures by the maximum-likelihood method. *Acta Crystallogr D Biol Crystallogr* 53(Pt 3):240–255.
8. Emsley P, Cowtan K (2004) Coot: model-building tools for molecular graphics. *Acta Crystallogr D Biol Crystallogr* 60(Pt 12 Pt 1):2126–2132.
9. Vagin A, Teplyakov A (2010) Molecular replacement with MOLREP. *Acta Crystallogr D Biol Crystallogr* 66(Pt 1):22–25.
10. Brennich ME, et al. (2016) Online data analysis at the ESRF bioSAXS beamline, BM29. *J Appl Crystallogr* 49:203–212.
11. Franke D, Jeffries CM, Svergun DI (2015) Correlation Map, a goodness-of-fit test for one-dimensional X-ray scattering spectra. *Nat Methods* 12(5):419–422.
12. Petoukhov MV, et al. (2012) New developments in the ATSAS program package for small-angle scattering data analysis. *J Appl Crystallogr* 45(Pt 2):342–350.
13. Guinier A (1939) *La Diffraction des rayons x aux très petits angles: application à l'étude des*

- phénomènes ultramicroscopiques. *Ann Phys* 12:161–237.
14. Porod G (1982) General theory. *Small-Angle X-Ray Scattering*, eds Glatter O, Kratky O (Academic Press, London).
 15. Rambo RP, Tainer JA (2013) Accurate assessment of mass, models and resolution by small-angle scattering. *Nature* 496(7446):477–481.
 16. Svergun DI, Petoukhov MV, Kochet MH (2001) Determination of domain structure of proteins from X-ray solution scattering. *Biophys J* 80(6):2946–2953.
 17. Volkov VV, Svergun DI (2003) Uniqueness of ab initio shape determination in small-angle scattering. *J Appl Crystallogr* 36:860–864.
 18. Taylor TC, Andersson I (1997) Structure of a product complex of spinach Ribulose-1,5-Bisphosphate Carboxylase/Oxygenase. *Biochemistry* 36(13):4041–4046.
 19. Fermani S, et al. (2010) The crystal structure of photosynthetic glyceraldehyde-3-phosphate dehydrogenase (isoform A4) from *Arabidopsis thaliana* in complex with NAD. *Acta Crystallogr Sect F Struct Biol Cryst Commun* 66(Pt6):621–626.
 20. Zaffagnini M, et al. (2014) High resolution crystal structure and redox properties of chloroplastic triosephosphate isomerase from *Chlamydomonas reinhardtii*. *Mol Plant* 7(1):101–120.
 21. Chiadmi M, Navaza A, Miginiac-Maslow M, Jacquot JP, Cherfils J (1999) Redox signalling in the chloroplast: Structure of oxidized pea fructose-1,6-bisphosphate phosphatase. *EMBO J* 18(23):6809–6815.
 22. Pasquini M, et al. (2017) Structural basis for the magnesium-dependent activation of transketolase from *Chlamydomonas reinhardtii*. *Biochim Biophys Acta* 1861(8):2132–2145.
 23. Gütle DD, et al. (2016) Chloroplast FBPase and SBPase are thioredoxin-linked enzymes with similar architecture but different evolutionary histories. *Proc Natl Acad Sci USA* 113(24):6779–6784.
 24. Kopp J, Kopriva S, Süß KH, Schulz GE (1999) Structure and mechanism of the amphibolic

- enzyme D-ribulose-5-phosphate 3-epimerase from potato chloroplasts. *J Mol Biol* 287(4):761-771.
25. Davies GJ, et al. (1994) Structure of the ADP complex of the 3-phosphoglycerate kinase from *Bacillus stearothermophilus* at 1.65 Å. *Acta Crystallogr D Biol Crystallogr* 50(Pt 2):202-209.
26. Heron PW, Sygusch J (2017) Isomer activation controls stereospecificity of class I fructose-1,6-bisphosphate aldolases. *J Biol Chem* 292(48):19849-19860.
27. Dereeper A, et al. (2008) Phylogeny.fr: robust phylogenetic analysis for the non-specialist. *Nucleic Acids Res* 36 (Web Server issue):W465-W469.
28. Charlier HA, Runquist JA, Miziorko HM (1994) Evidence supporting catalytic roles for aspartate residues in phosphoribulokinase. *Biochemistry* 33(31):9343–9350.
29. Thieulin-Pardo G, Remy T, Lignon S, Lebrun R, Gontero B (2015) Phosphoribulokinase from *Chlamydomonas reinhardtii*: a Benson-Calvin cycle enzyme enslaved to its cysteine residues. *Mol. Biosyst.* 11(4):1134–1145.
30. Letunic I, Bork P (2016) Interactive tree of life (iTOL) v3: an online tool for the display and annotation of phylogenetic and other trees. *Nucleic Acids Res* 44(W1):242-245.
31. Harrison DH, Runquist JA, Holub A, Miziorko HM (1998) The crystal structure of phosphoribulokinase from *Rhodobacter sphaeroides* reveals a fold similar to that of adenylate kinase. *Biochemistry* 37(15):5074–5085.
32. Robert X, Gouet P (2014) Deciphering key features in protein structures with the new ENDscript server. *Nucleic Acids Res* 42:W320-W324.
33. Sievers F, et al. (2011) Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Mol Syst Biol* 7:539-545.
34. Capitani G, et al. (2000) Crystal structures of two functionally different thioredoxins in spinach chloroplasts. *J Mol Biol* 302(1):135-154.
35. Bienert S, et al. (2017) The SWISS-MODEL Repository - new features and functionality.

- Nucleic Acids Res 45(D1):D313-D319.
36. Hallenbeck PL, Kaplan S (1987) Cloning of the gene for phosphoribulokinase activity from *Rhodobacter sphaeroides* and its expression in *Escherichia coli*. *J Bacteriol* 169(8):3669-3678.
 37. Meijer WG, et al. (1991) Identification and organization of carbon dioxide fixation genes in *Xanthobacter flavus* H4-14. *Mol Gen Genet* 225(2):320-330.
 38. Kobayashi D, Tamoi M, Iwaki T, Shigeoka S, Wadano A (2003) Molecular characterization and redox regulation of phosphoribulokinase from the cyanobacterium *Synechococcus* sp. PCC 7942. *Plant Cell Physiol* 44(3):269–276.
 39. Kossmann J, Klintworth R, Bowien B (1989) Sequence analysis of the chromosomal and plasmid genes encoding phosphoribulokinase from *Alcaligenes eutrophus*. *Gene* 85(1):247-252.
 40. Roesler KR, Ogren WL (1990) *Chlamydomonas reinhardtii* phosphoribulokinase: sequence, purification and kinetics. *Plant Physiol* 93(1):188–193.
 41. Michalowski CB, Derocher EJ, Bohnert HJ, Salvucci ME (1992) Phosphoribulokinase from ice plant: Transcription, transcripts and protein expression during environmental stress. *Photosynth Res* 31(2):27-38.
 42. Burki F, et al. (2010) Evolution of Rhizaria: new insights from phylogenomic analysis of uncultivated protists. *BMC Evol Biol* 10:377-395.
 43. Okamoto K, Kondo-Okamoto N, Ohsumi Y (2009) A landmark protein essential for mitophagy. *Autophagy* 5(8):1203-1205.