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Trapping membrane proteins with amphipols.
Structure and properties of membrane protein/amphipol complexes

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1) How to trap a membrane protein (MP) in amphipols (APols)?

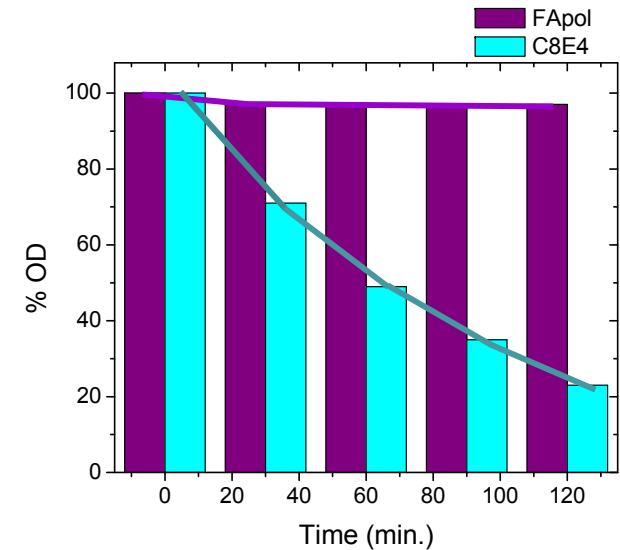
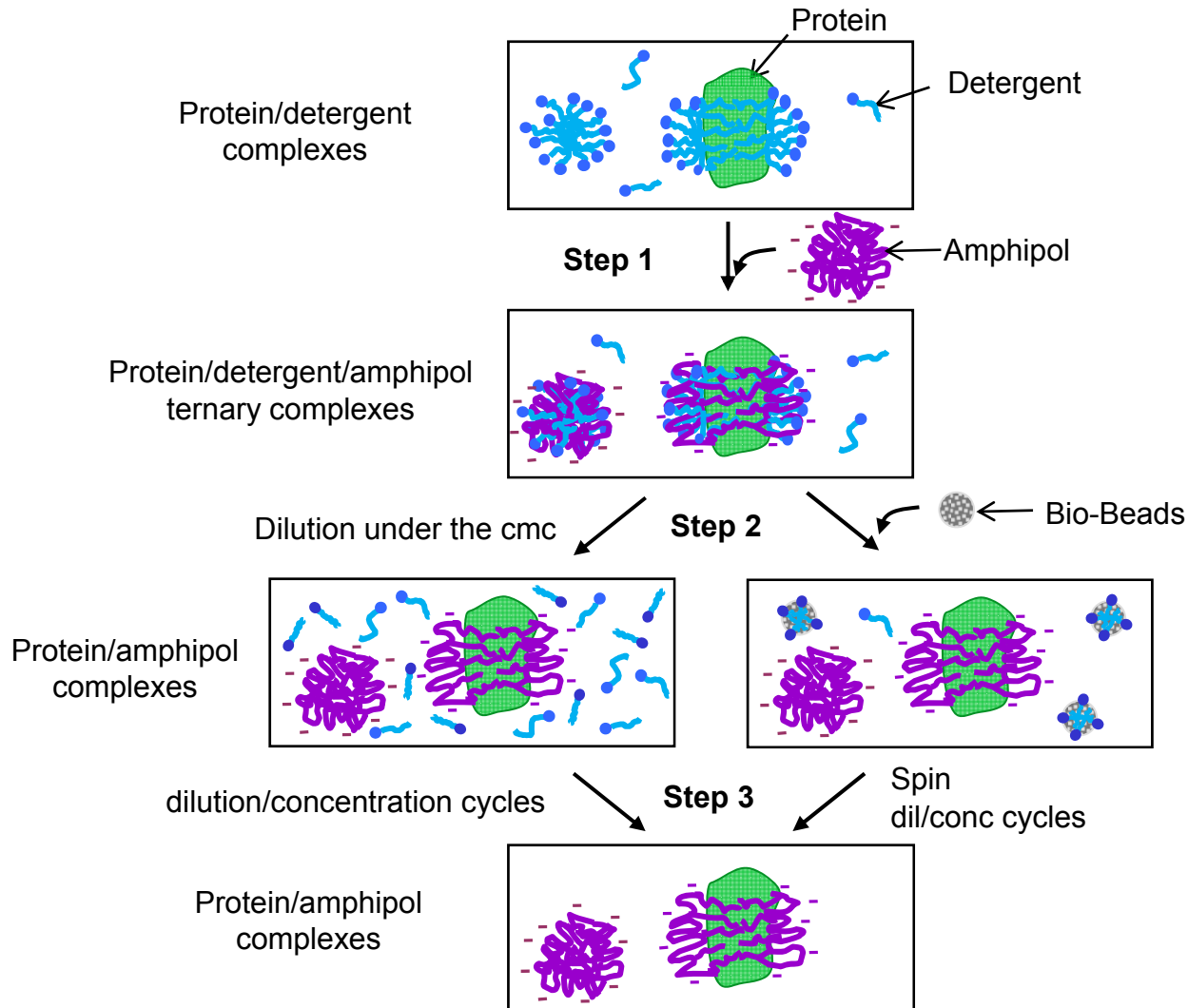
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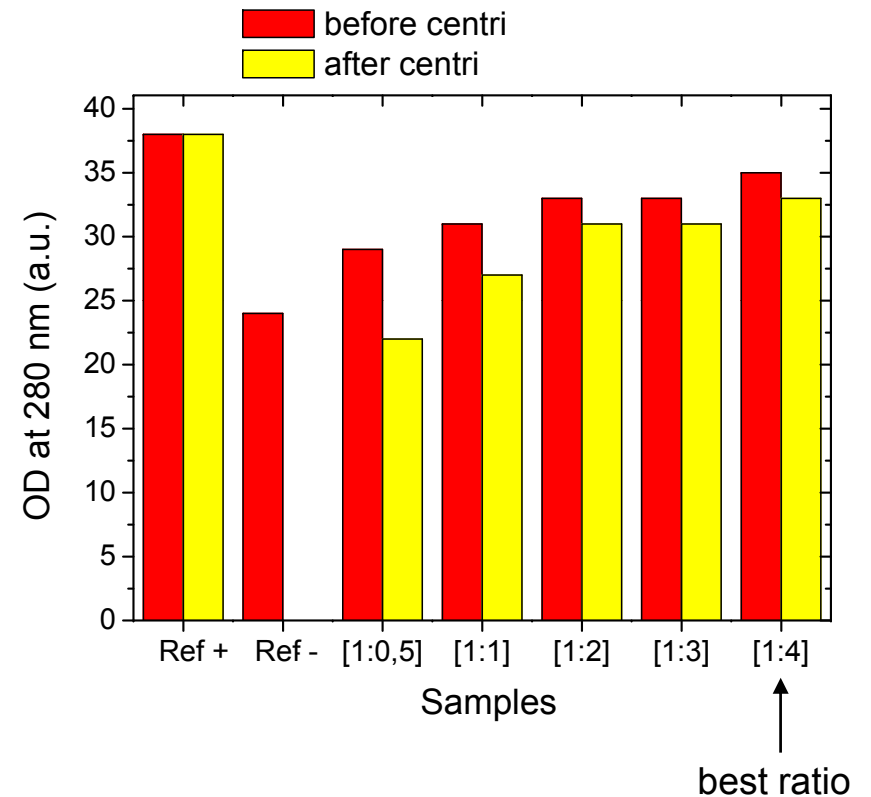
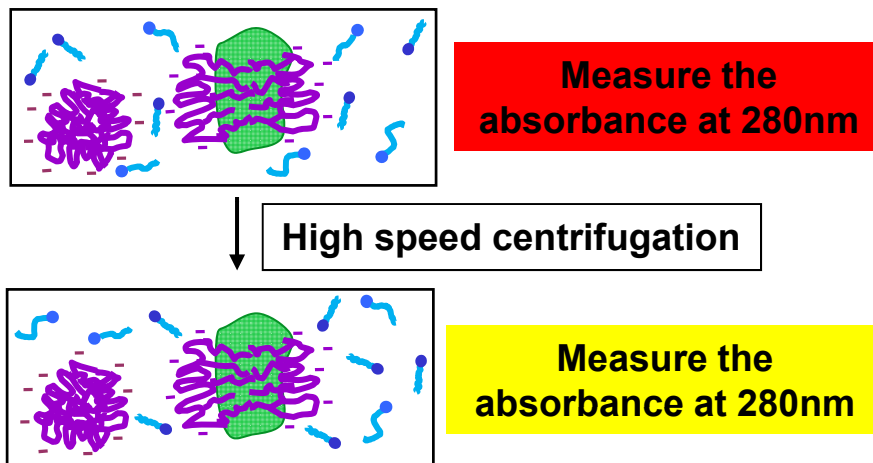
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1) How to define conditions for trapping a MP in APols?

- Estimation of the best [protein:APol] w/w ratio

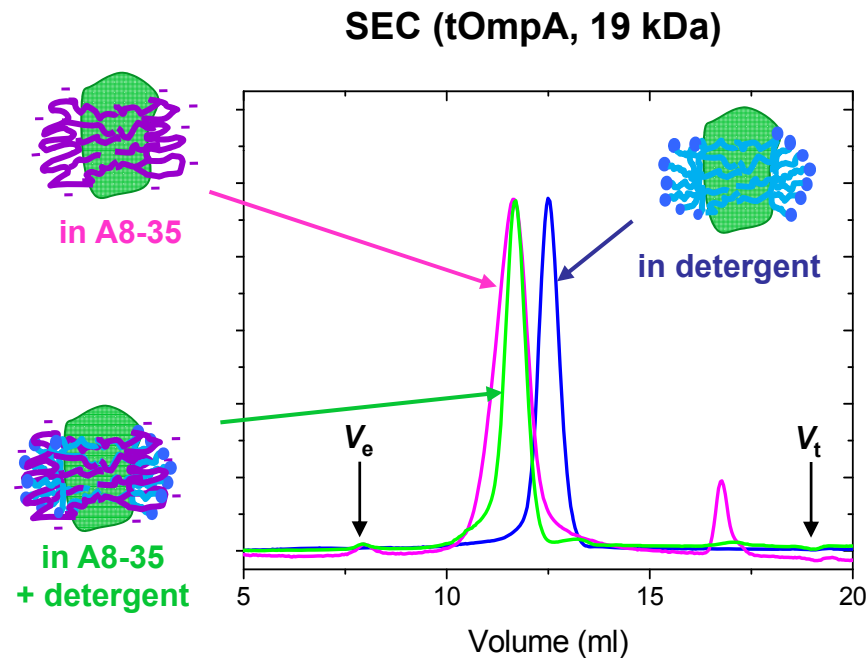
Trapping after dilution under the cmc



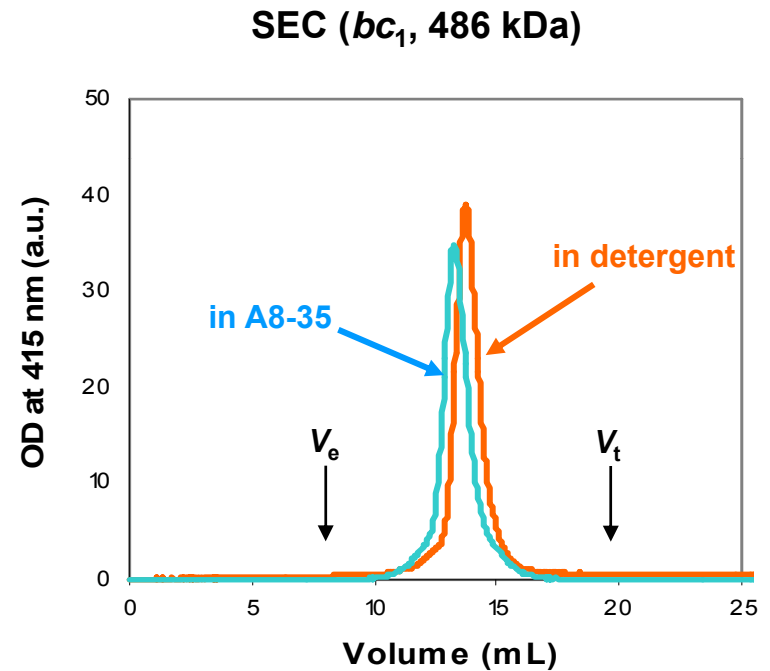
- Analyze the resulting complexes by size exclusion chromatography (SEC)

2) What do MP/APol complexes look like after trapping?

1. MP/APol complexes are slightly larger than MP/detergent complexes (SEC, SANS, AUC, NMR)



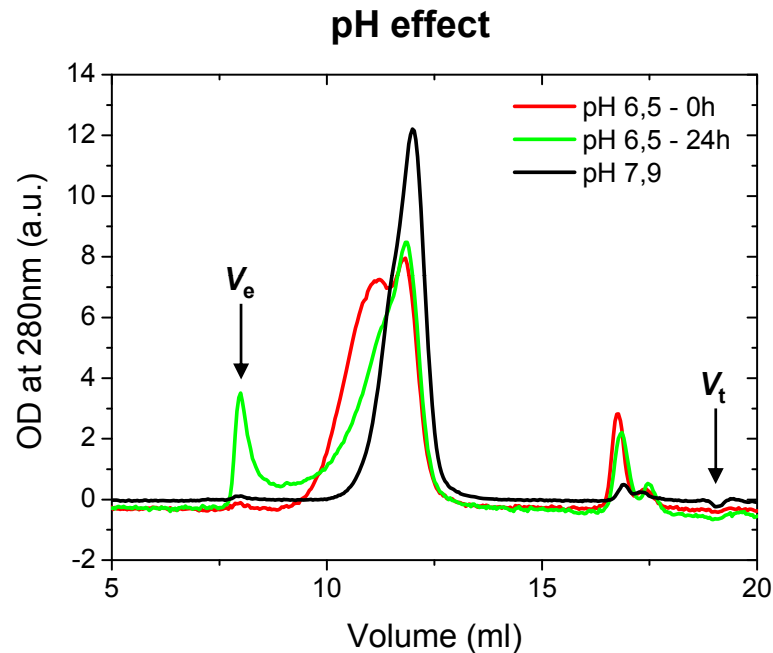
Zoonens *et al.*, *Biochemistry*, 2007



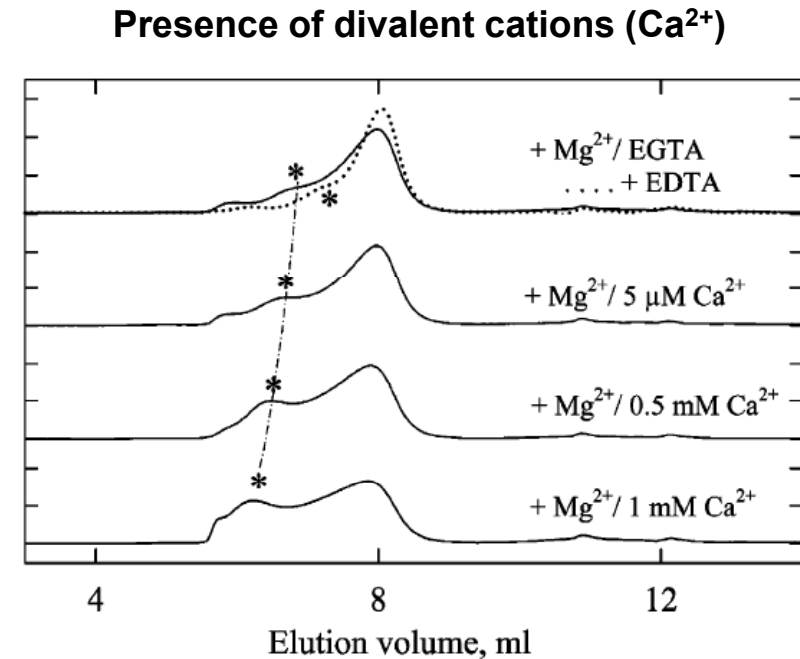
Charvolin *et al.*, *in preparation*

2) What do MP/APol complexes look like after trapping?

1. MP/APol complexes are slightly larger than MP/detergent complexes
2. The monodispersity of the complexes depends on several factors



Zoonens *et al.*, *Biochemistry*, 2007



Picard *et al.*, *Biochemistry*, 2006

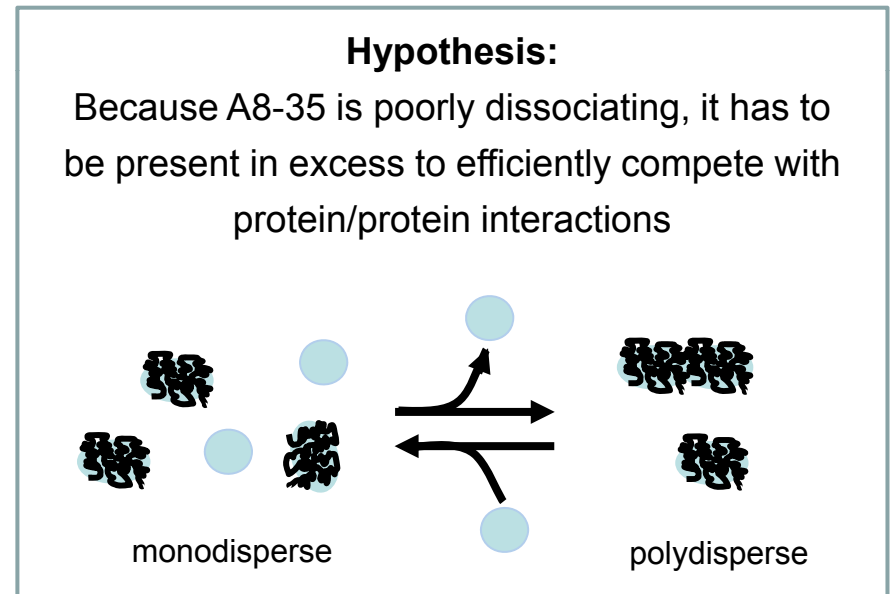
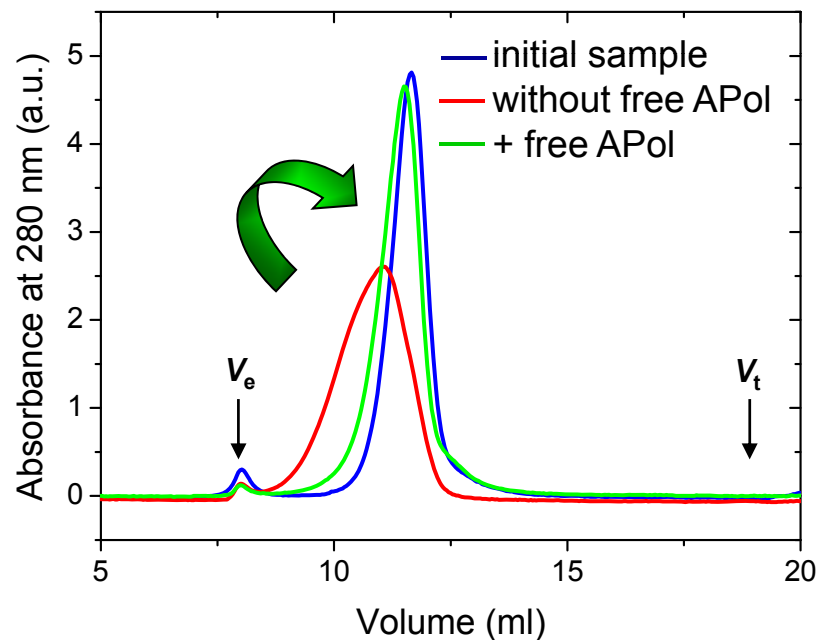
Hypothesis:

Because the solubility of A8-35 is conferred by its charges, lowering pH or the presence of divalent cations reduces the electrostatic repulsions between particles. Divalent cations could also link up particles together and lead to aggregation.

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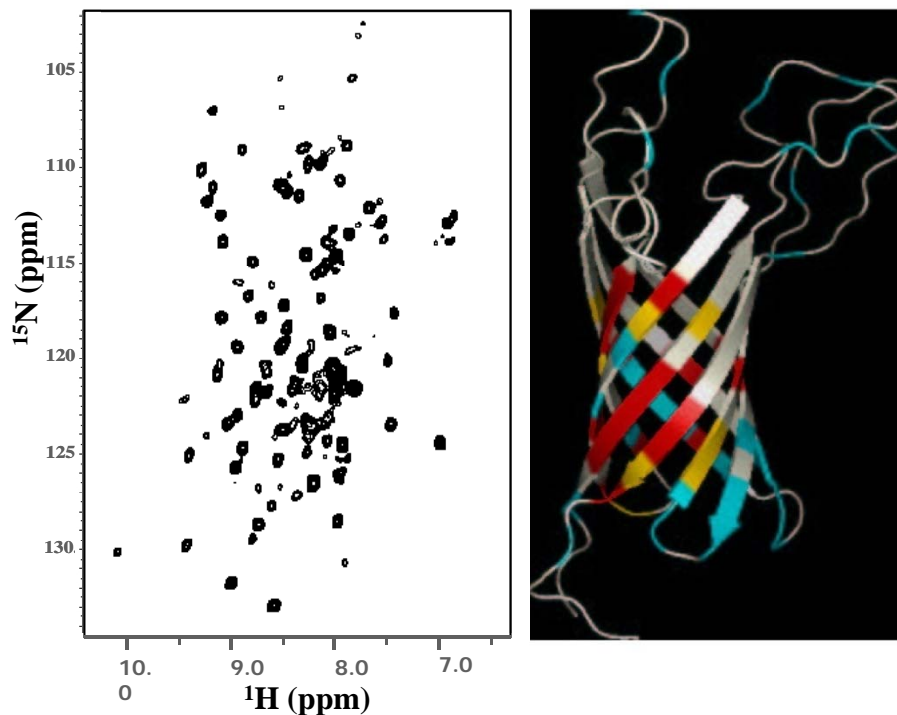
Separating the complexes from extra free APol



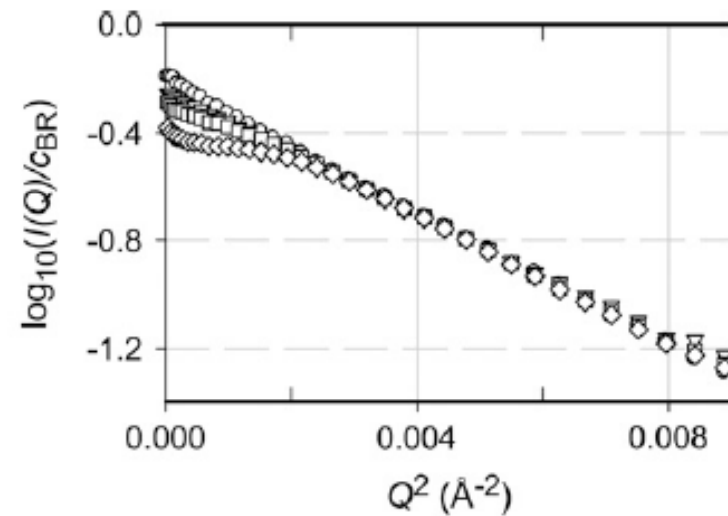
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2. The monodispersity of the complexes depends on several factors
3. Amphipols form a **compact layer** (1.5-2 nm) around the **transmembrane surface** of the protein (no diffuse corona; SANS, NMR, AUC)

NMR (tOmpA, 19 kDa)



SANS (BR, 27 kDa)



Gohon *et al.*, *Biophys. J.*, 2008

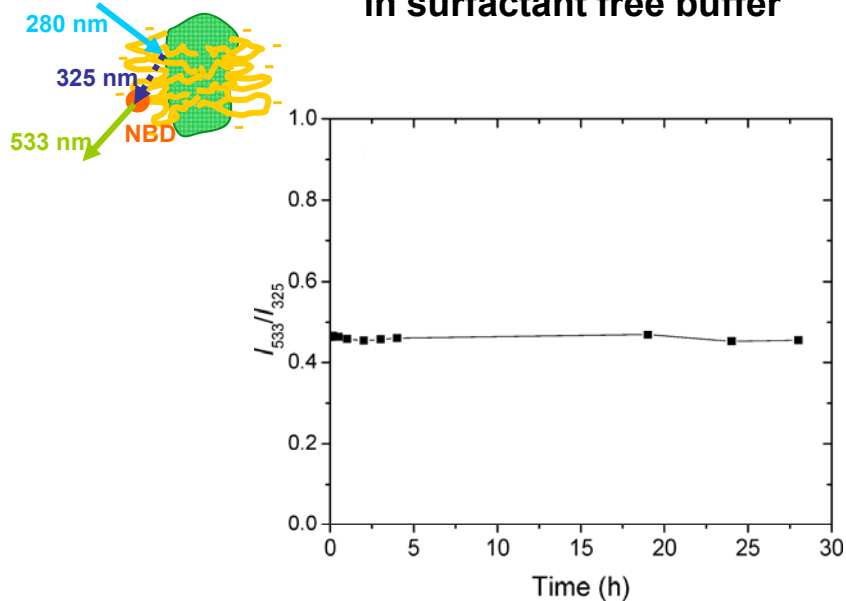
Zoonens *et al.*, *PNAS*, 2005

Catoire *et al.*, *Eur Biophys J*, 2010

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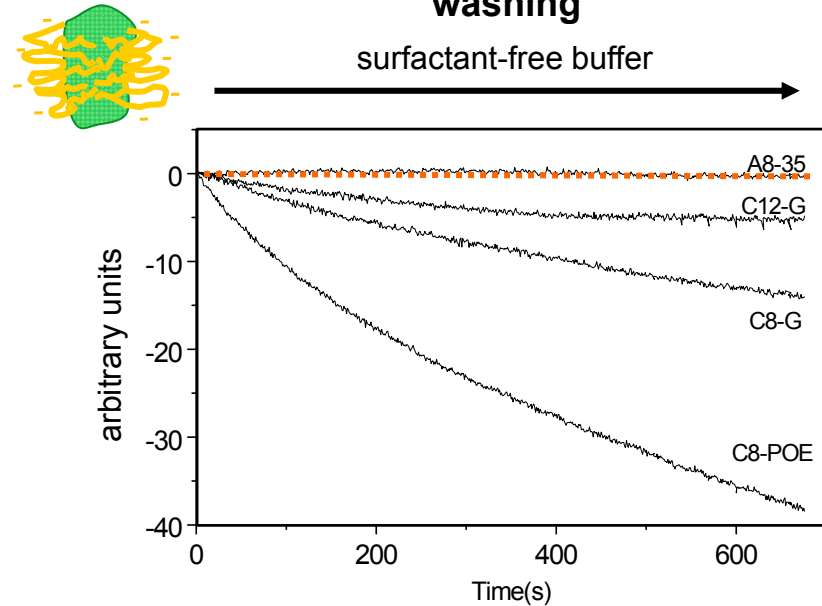
1. MP/APol complexes are slightly larger than MP/detergent complexes
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3. Amphipols form a compact layer around the transmembrane surface of the protein
4. Binding is non-covalent, but irreversible in the absence of a competing surfactant

**FRET measurements
after 1000-fold dilution
in surfactant free buffer**



Zoonens *et al.*, *Biochemistry*, 2007

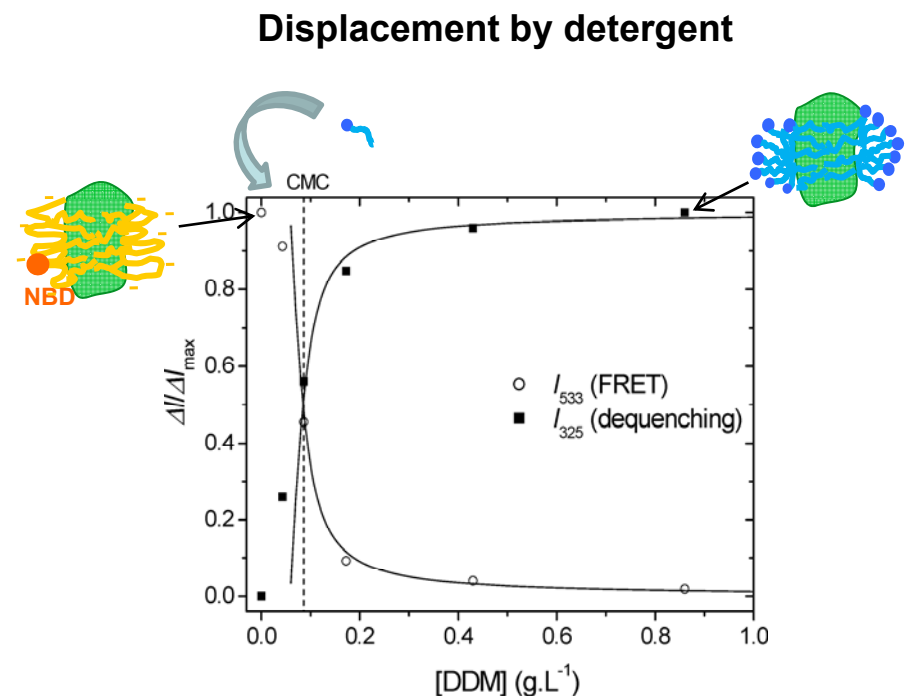
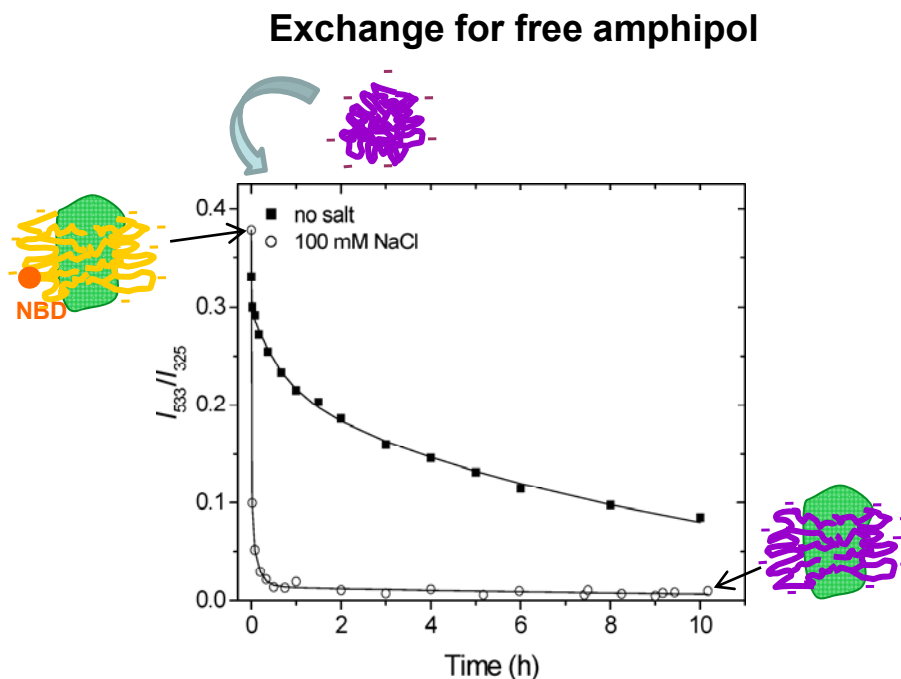
**Surface Plasmon Resonance
measurements upon extensive
washing**



Hong & Lakey; *in* Popot *et al.*, *CMLS*, 2003

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3. Amphipols form a compact layer around the transmembrane surface of the protein
4. Binding is non-covalent, but irreversible in the absence of a competing surfactant
5. Bound amphipols can be displaced by free amphipols, detergents, or lipids (Tribet *et al.*, *Langmuir*, 1997; Nagy *et al.*, *FEBS Lett.*, 2001; Pocanschi *et al.*, *Biochemistry*, 2006; Zoonens *et al.*, *Biochemistry*, 2007; Tribet *et al.*, *Langmuir*, 2009)

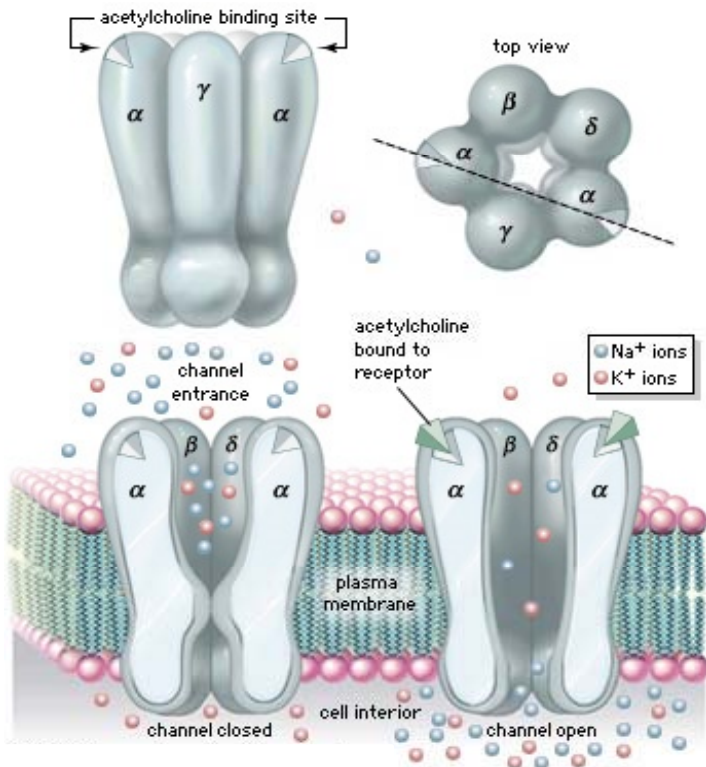


Zoonens *et al.*, *Biochemistry*, 2007

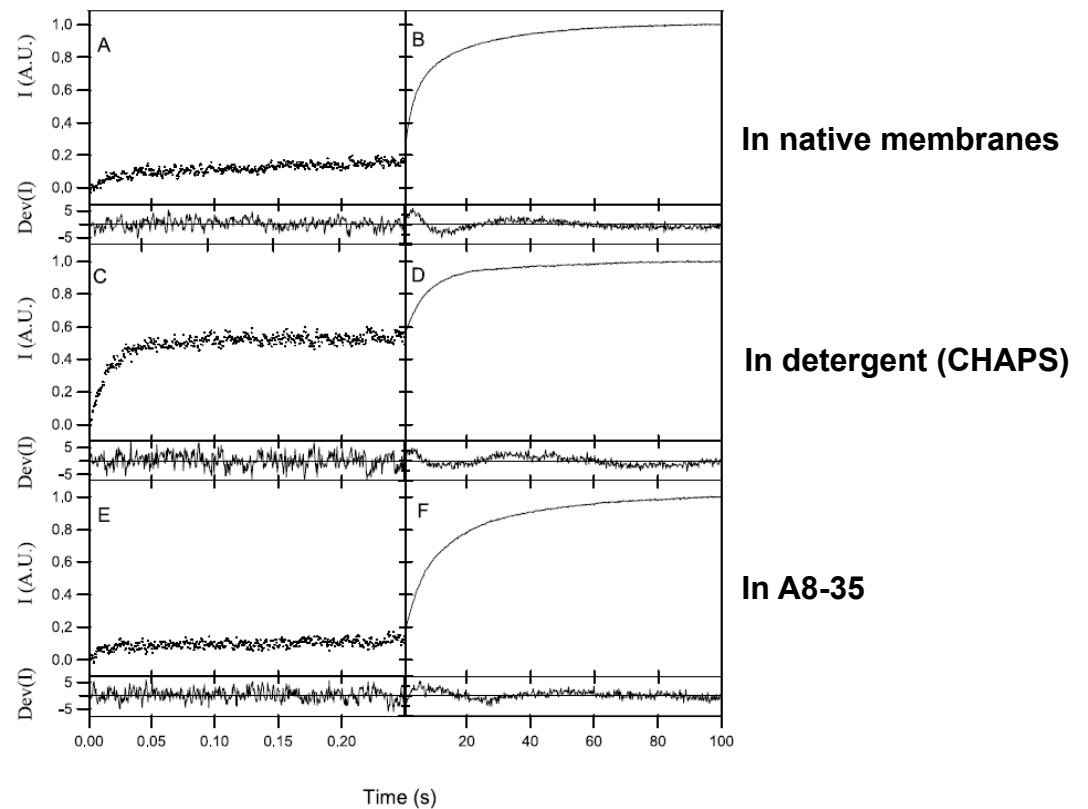
3) How do MP/APol complexes behave in term of activity ?

1. Amphipols may affect the dynamics –and, thereby, the activity– of the proteins they bind to

- Nicotinic acetylcholine receptor: no transmembrane movement; allosteric transitions unaffected



Kinetics of binding

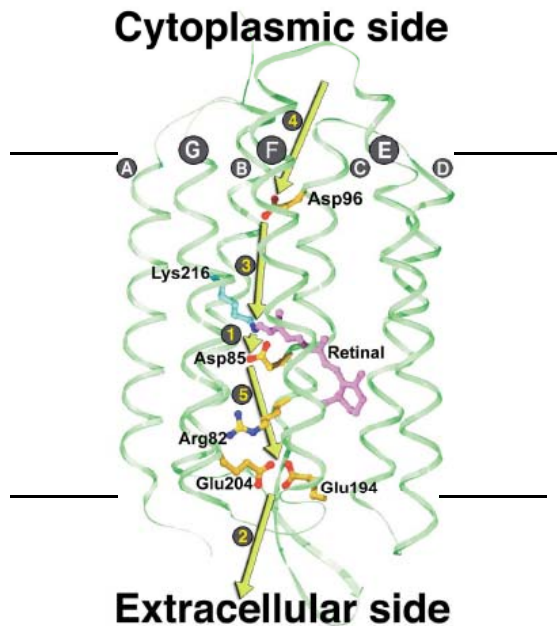


Martinez *et al.*, *FEBS Lett.*, 2002

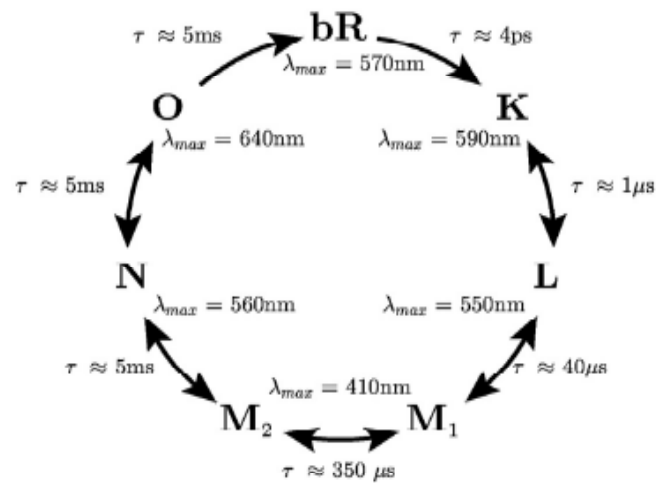
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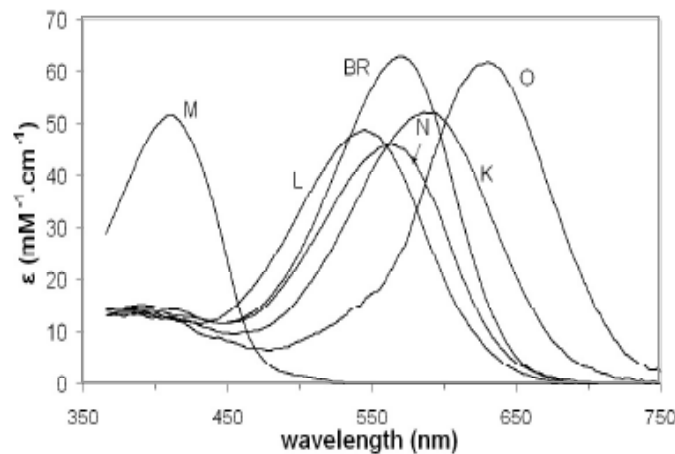
- Nicotinic acetylcholine receptor: no transmembrane movement; allosteric transitions unaffected
- Bacteriorhodopsin: very small transmembrane movements; no or very limited effects on the photocycle



Neutze *et al.*, *BBA*, 2002



Spectral changes

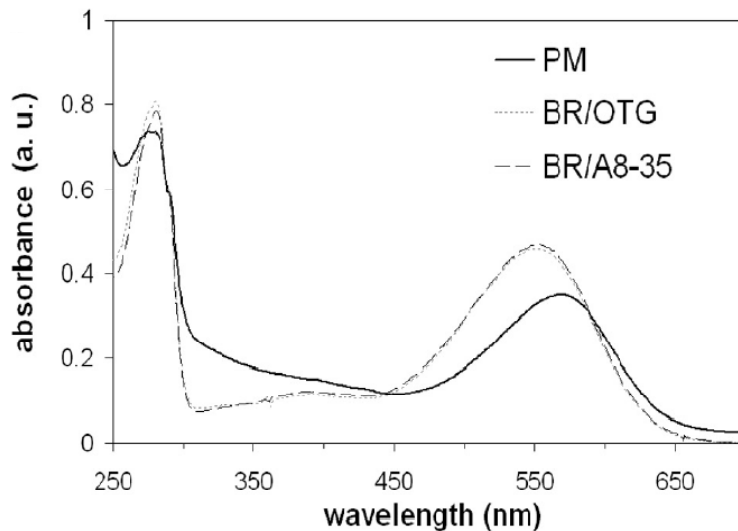


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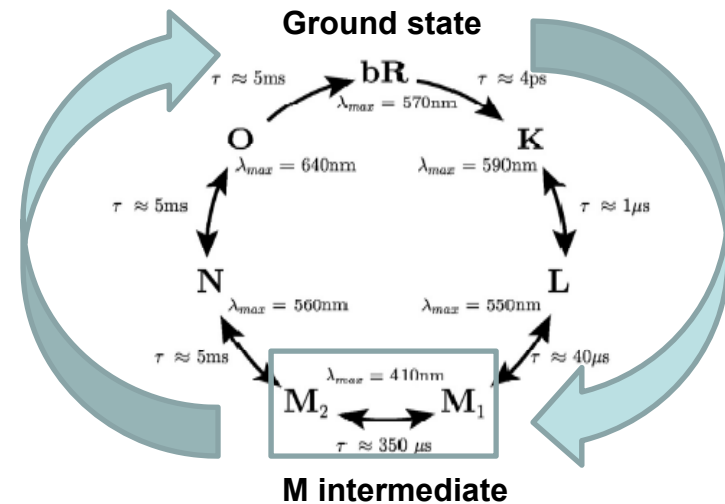
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UV-visible spectra of BR



Gohon *et al.*, *Biophys. J.*, 2008



	DAS1 (ns)	DAS2 (μs)	DAS3 (μs)	DAS4 (ms)	DAS5 (ms)
PM	650	21	84	1.4	4.7
BR/OTG	375	4.3	23	0.53	9
BR/A8-35	480	5.8	53	1.0	6.3

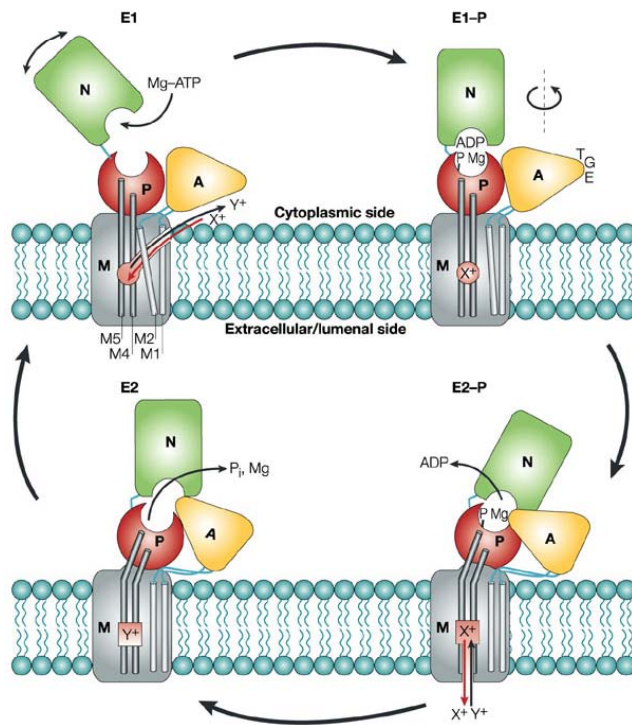
M's rise

M's decay

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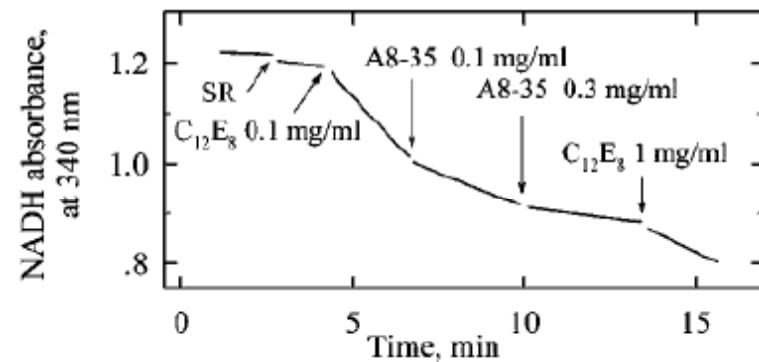
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- Sarcoplasmic calcium ATPase: large-scale transmembrane rearrangements; ATP hydrolysis and Ca^{2+} release reversibly inhibited Champeil et al., JBC 2000; Picard et al., *Biochemistry* 2006

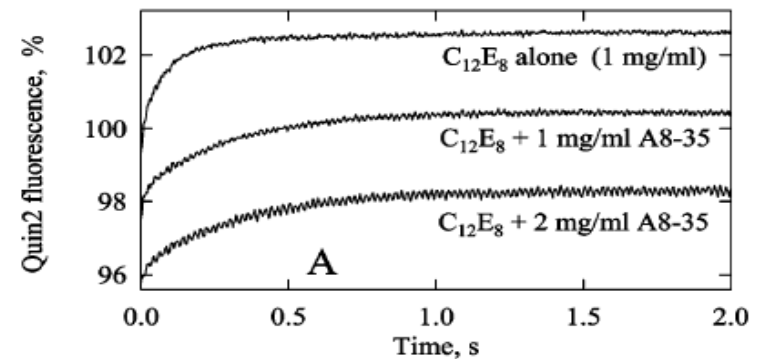


Kühlbrandt, *Nature Reviews Molecular Cell Biology*, 2004

Hydrolytic activity of Ca^{2+} ATPase



Ca^{2+} dissociation experiment

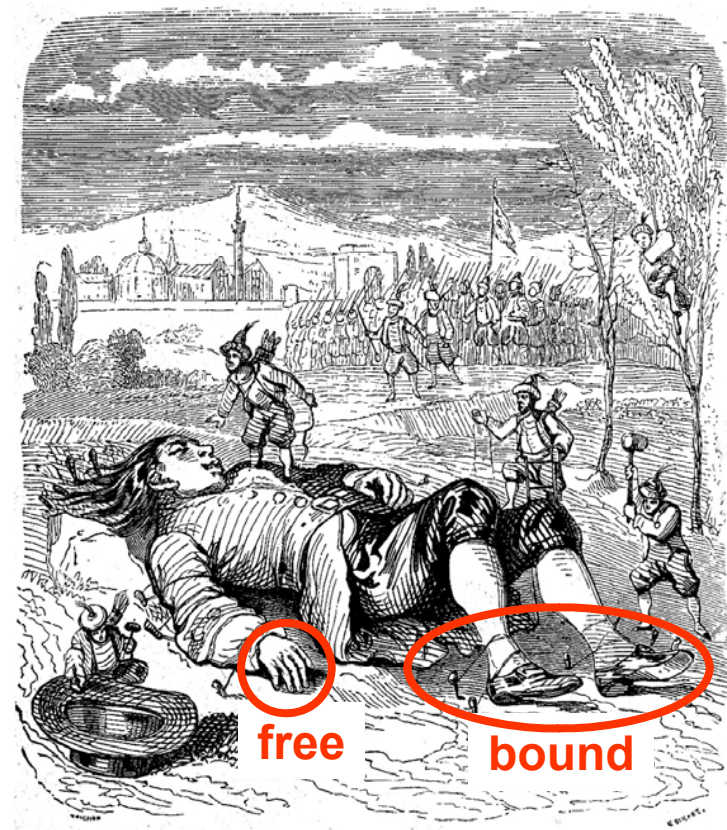


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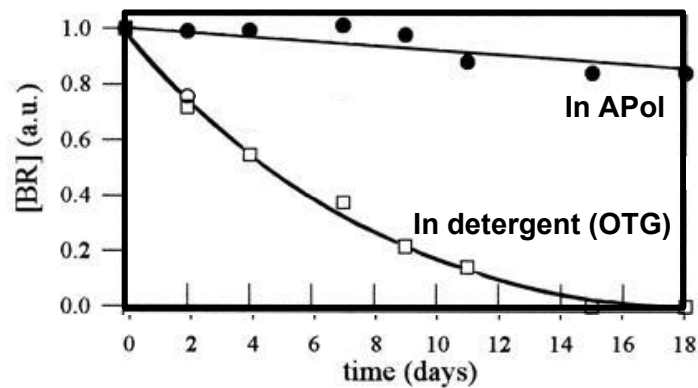
⇒ damping of large-scale transmembrane movements ('Gulliver effect')



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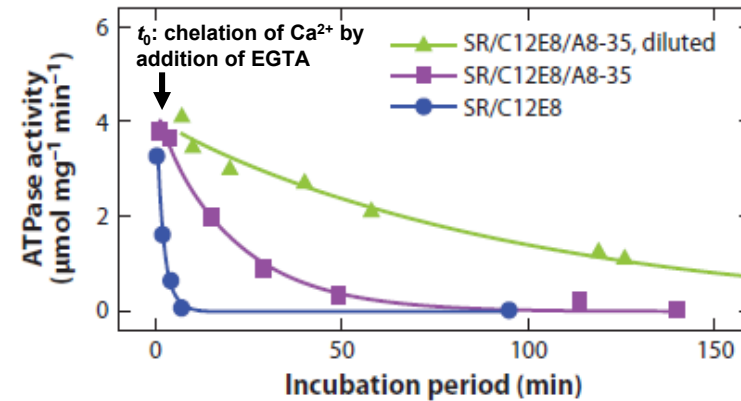
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2. Damping of dynamics may contribute to membrane protein stabilization by amphipols

Bacteriorhodopsin



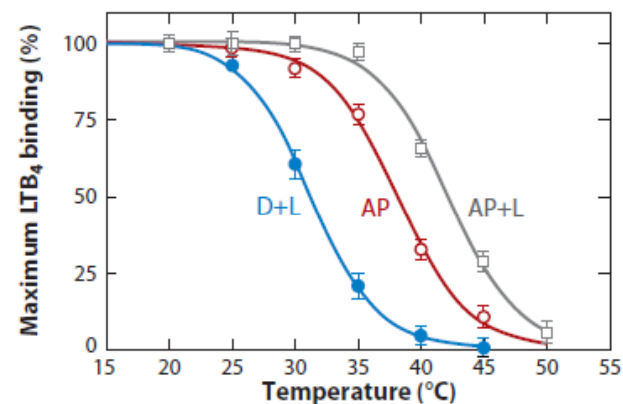
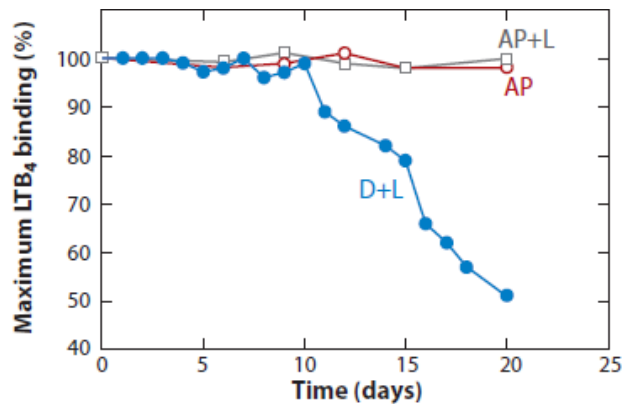
Tribet *et al.*, (1996) *PNAS* **93**, 15047

Calcium ATPase



Champeil *et al.*, *JBC* 2000

Leukotriene receptor BLT1 (GPCR)



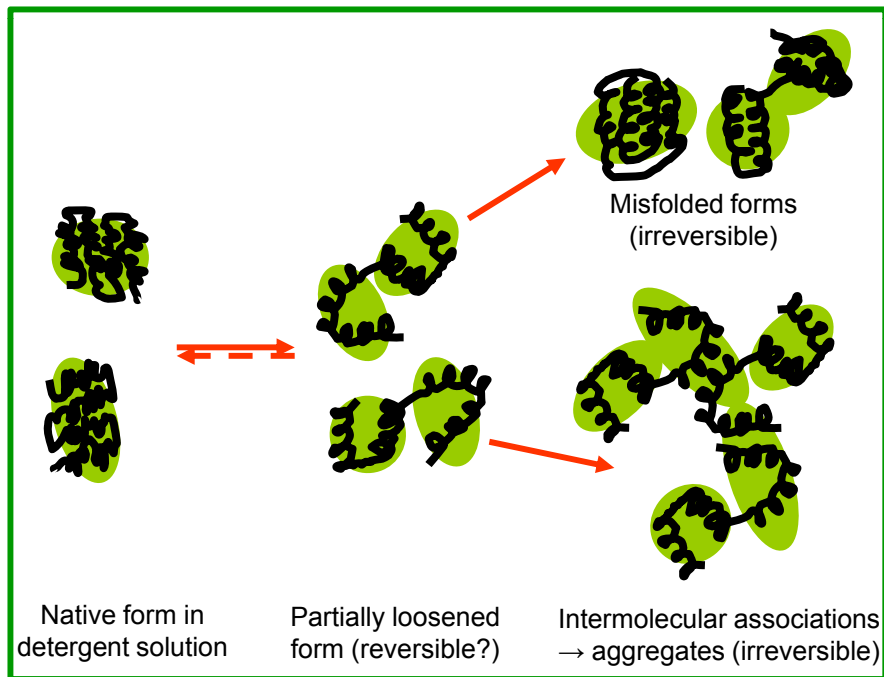
Dahmane *et al.*,
Biochemistry, 2009

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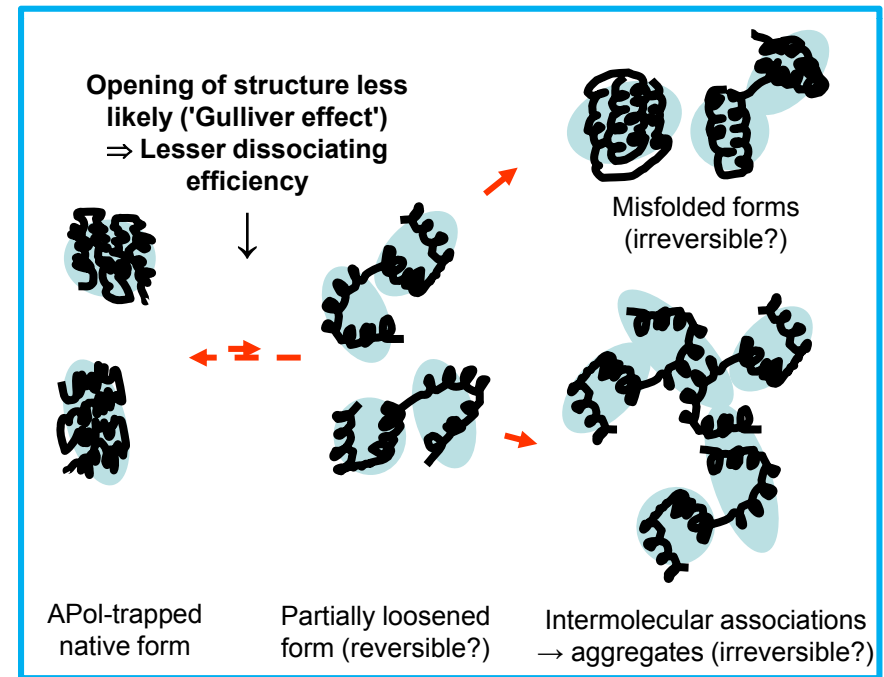
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⇒ Could the 'Gulliver effect' contribute to stabilizing APol-trapped MPs against inactivation ?

Denaturation by detergents



Stabilization by amphipols



For discussion, see Popot *et al.*, *CMLS*, 2003; Picard *et al.*, *Biochemistry*, 2006.

In conclusion

- Protocol of trapping: Usually, we transfer MPs from detergent solution to APols
- Solution properties: MP/APol complexes are essentially homogeneous but the monodispersity depends on the pH (higher than 7), the absence of divalent cations, and the presence of extra free APols
- Structural organization: APols interact exclusively with the hydrophobic transmembrane surface of MPs and form a compact layer of 1.5 to 2nm thickness
- Dynamics of association: APols do not desorb from MPs but they exchange for other surfactants (detergent, APols, or lipids)
- Activity: It seems to depend on the amplitude of the transmembrane conformational changes occurring during the catalytic cycle of the protein of interest
- Stability: MPs trapped in APols are generally more stable than in detergent solution
- Ligand binding: Generally unaffected by Apol trapping